

Bioremediation: the Hope and the Hype

*biological processes for environmental
clean-up (good, bad, and ugly)*



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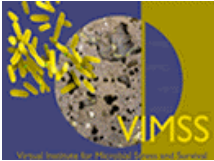
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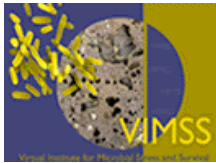
The Problem

73,000,000 U.S.
Citizens

live within 4 miles of
Superfund Site

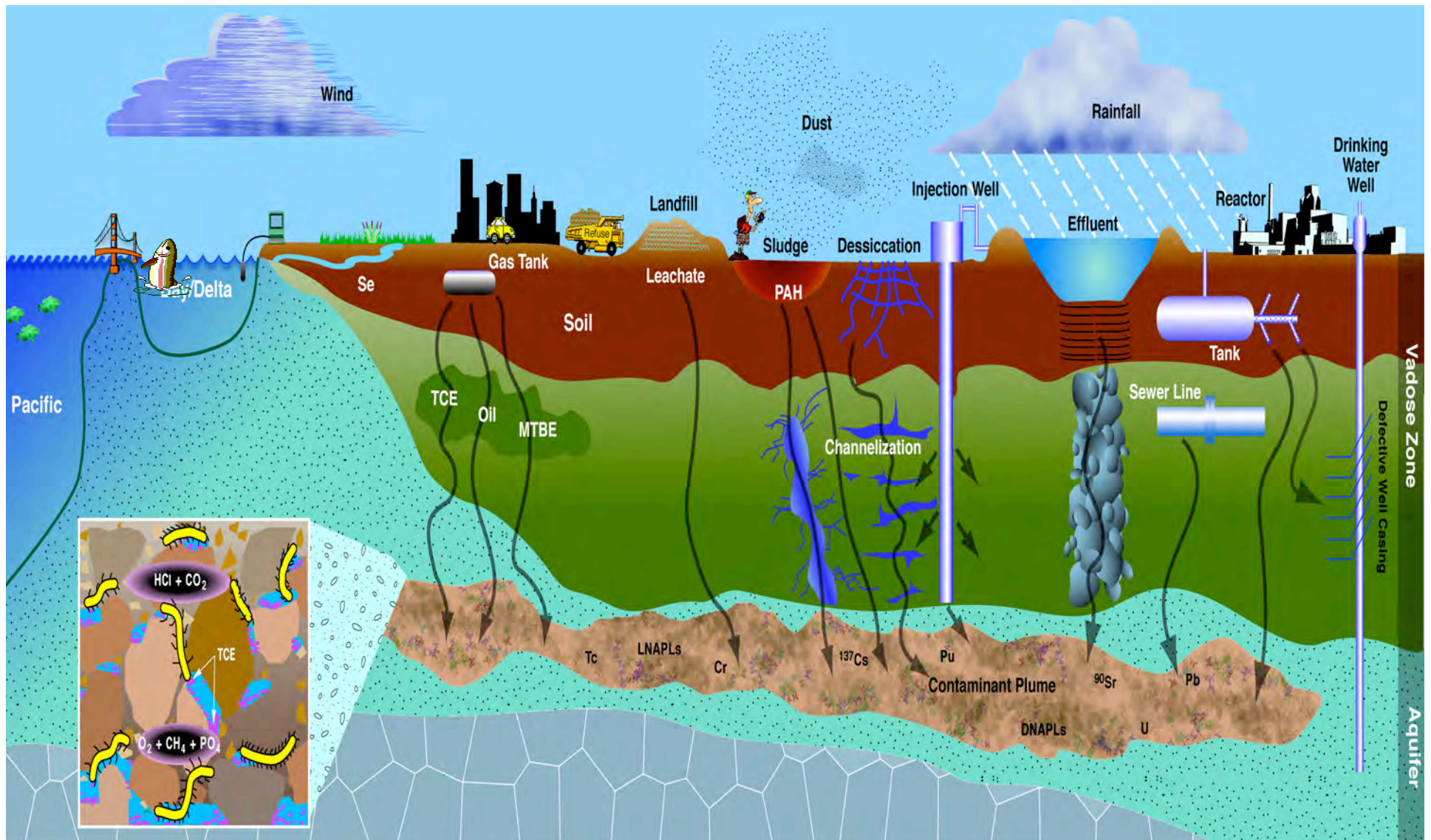
4,000,000 U.S. Citizens
live within 1 mile of
a Superfund Site

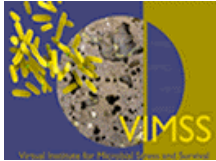




The Sources

Understanding, monitoring and controlling the environment
(the need for biosustainability everywhere)





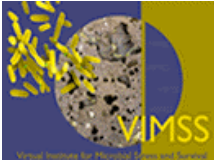
The Cost



Hazardous Waste Remediation
in the United States
could cost



> \$1.7 Trillion

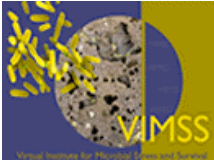


Amoco Cadiz Spill 1978

- 227,000 tons heavy crude oil
- Entire Normandy coast impacted
- Spill was so large they only treated areas that impacted economy the most
- Large areas in remote parts of coast abandoned
- Best available treatment - detergents (dispersants)

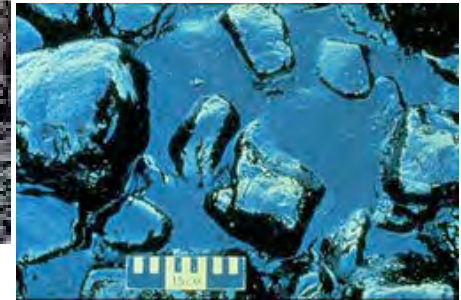


1. Spill was impetus for variety of international cleanup and tanker regulations.
2. Ecological studies done 10 and 20 yrs after the spill have demonstrated that
 - the areas that were *'treated'* have not recovered yet
 - the areas that were *untreated* recovered in < 5 yr!!!!!!

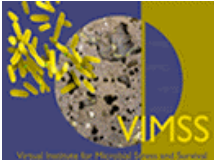


18 yrs later Exxon Valdez spill

- 3/24/89 Supertanker spills 11 million gallons of crude into Prince William Sound, 1,300 miles of coastline impacted (largest spill in US history)
- Cleanup involved: burning, mechanical, dispersants, and bioremediation
- Litigation is still going after 18 years, cost so far >\$7 billion

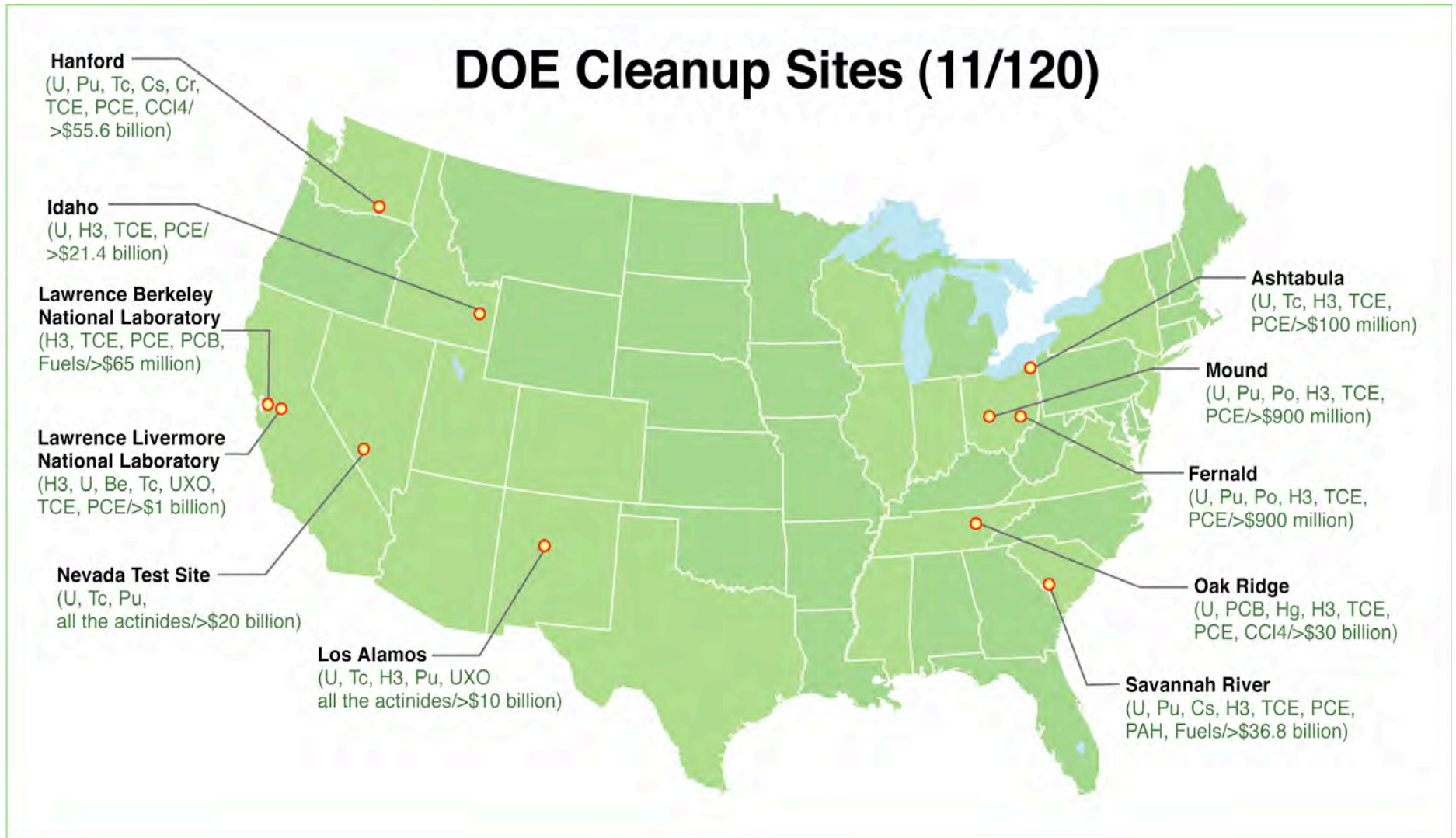


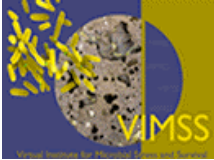
- Congress passed the 1990 Oil Pollution Act
- Fertilized areas were dramatically cleaner the first year, but no difference after the second winter
- Long term impact of treatments (dispersants and bioremediation) will severely impact ecology of sound for many decades



The DOE Problem

DOE Cleanup Sites (11/120)





Fernald, Ohio

Original Mission: Uranium purification 1952-1989, 500 million pounds, 1,050 acres near Cincinnati

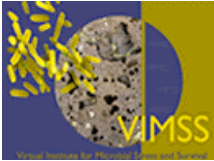
31 million pounds of nuclear product, 2.5 billion pounds of low-level radioactive, hazardous and mixed waste, and 2.5 million yd³ contaminated soil and debris.

Closed in 2006, ***“Weapons to Wetlands”!!!!!!!***

On-site Disposal Facility - 2.88 million yd³

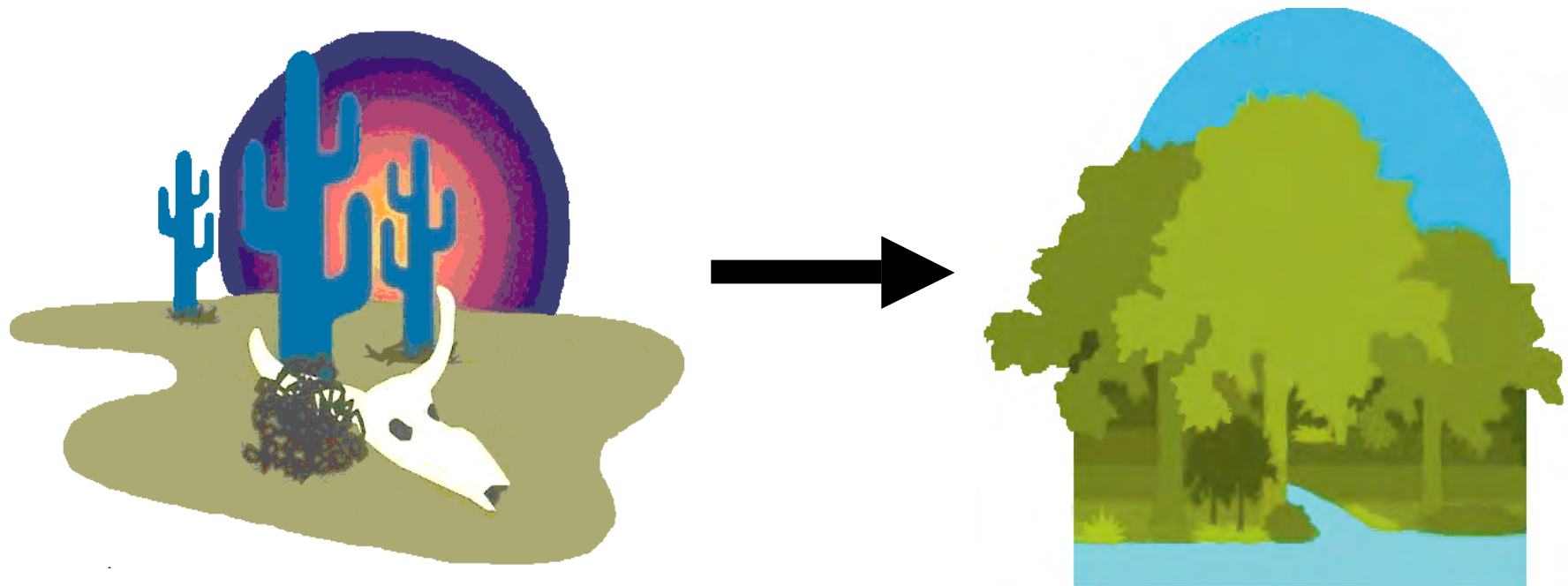
3 trains with 60 cars operating 24/7 hauling contaminated soil to EnviroCare, UT and Nevada Test Site for 5 years.

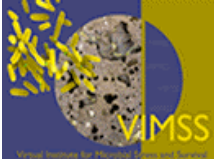




Bioremediation

Understanding, monitoring and controlling the environment for remediation with biological processes



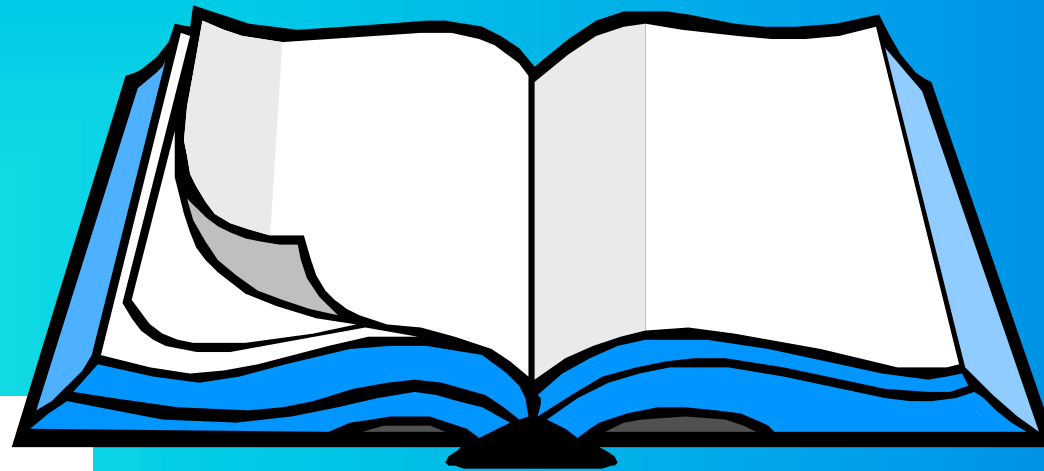


Benefits of Bioremediation

- Terminal Destruction
- On Site
- Environmentally Sound
- Cost Effective

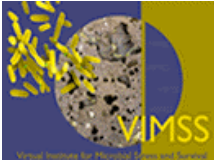
Bioremediation

First mention of
“bioremediation” in Scientific
Citation Index 1974–1996 in
abstract, title or keywords is in
1987.



Doctrine of Infallibility

There is no compound known to man,
man-made or natural,
that microorganisms can not degrade



Microbial* Life on Earth

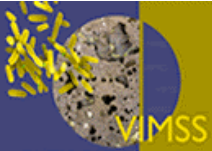
Cells

- 👉 Open Ocean 1.2×10^{29}
- 👉 Soil 2.6×10^{29}
- 👉 Oceanic Subsurface 3.5×10^{30}
- 👉 Terrestrial Subsurface $0.25-2.5 \times 10^{30}$
- 👉 All sources $4-6 \times 10^{30}$
- 👉 60% of all biomass on earth
- 👉 350-550 Pg of Carbon (60-100% more C than all plants)
- 👉 85-130 Pg of N and 9-14 Pg of P (10 times more than all plants)
- 👉 10^5-10^7 species
- 👉 Capable of 4 simultaneous mutations in every gene in 0.4 h
- 👉 Capable of dividing every 20 minutes
- 👉 Human Body 10^{14} cells with 10^{15} bacteria, 5K-10K species
- 👉 > 3.7 billion years of microbial evolution on earth



*** Prokaryotes only, Pg = 10^{15} g**

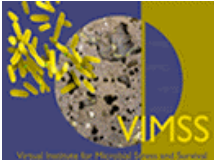
(in part Whitman et al., 1998)



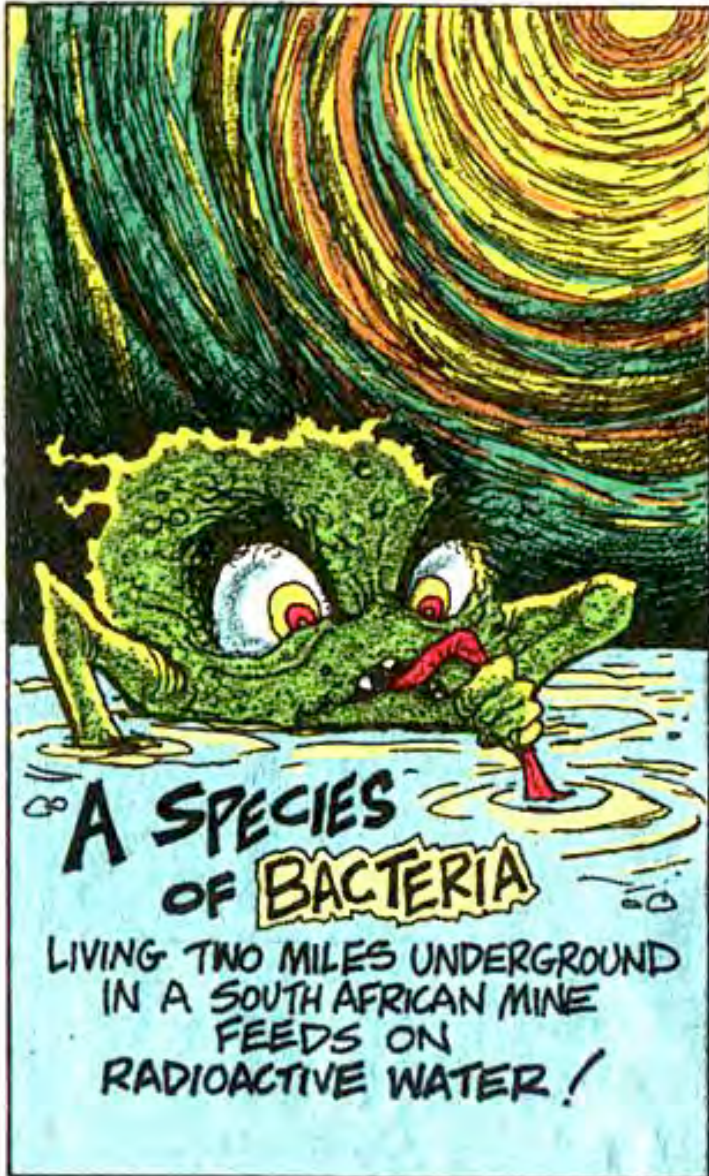
Microbial Growth Capabilities

<u>Factor</u>	<u>Lower Limit</u>	<u>Upper Limit</u>
Temperature	-12°C	104°C at 1000 ATM (sulfate reducing & oxidizing bacteria)
Eh	-400 to -450 mv at pH 8	+850 mv at pH 3 (CH ₄ producing bacteria)
pH	0 to 0.5 <i>Thiobacillus thiooxidans</i>	>13 <i>Plectonema sp.</i>
Hydrostatic Pressure	0	1400 ATM (deep sea bacteria)
Salinity	Double Distilled H ₂ O	Saturated Brines (halophilic bacteria)
Heavy Metals	<0.01 ppb	20,000 ppm Hg
Gases	CO ₂ , N ₂ , CH ₄ , H ₂ S, H ₂	

Note: These are conditions where microbes can grow, survival conditions are even more extreme.



Ripley's **Believe It or Not!**



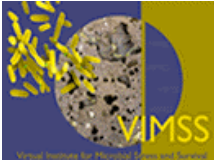
Lin et al. Science 10/06

Life in the slow lane.

- DNA was extracted from:
 - 3 Myr old fracture water
 - 8,000 ft. South Africa Gold Mine
 - Analogs to Mars?



**Homestake Mine
8,000 ft Lead, SD**



Normal Microbial Requirements

Parameters

Available soil water

Oxygen

Redox potential

pH

Nutrients

Temperature

Optimum Levels

25-85% holding capacity

Aerobic >0.2 mg/l DO

Anaerobic: $O_2 < 1\%$

Aerobes & Facultatives: > 50mv

Anaerobes: < 50mv

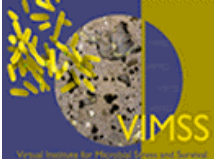
5.5-8.5

C:N:P of 120:10:1

15-45° C (Mesophiles)

“...1 g of soil typically contains 1 million to 10 billion microbial cells representing about 4,000-10,000 species...” (Torsvik et al., 1990)





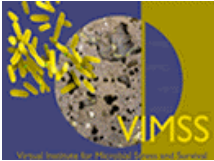
Factors that Affect Biodegradation

The Contaminant

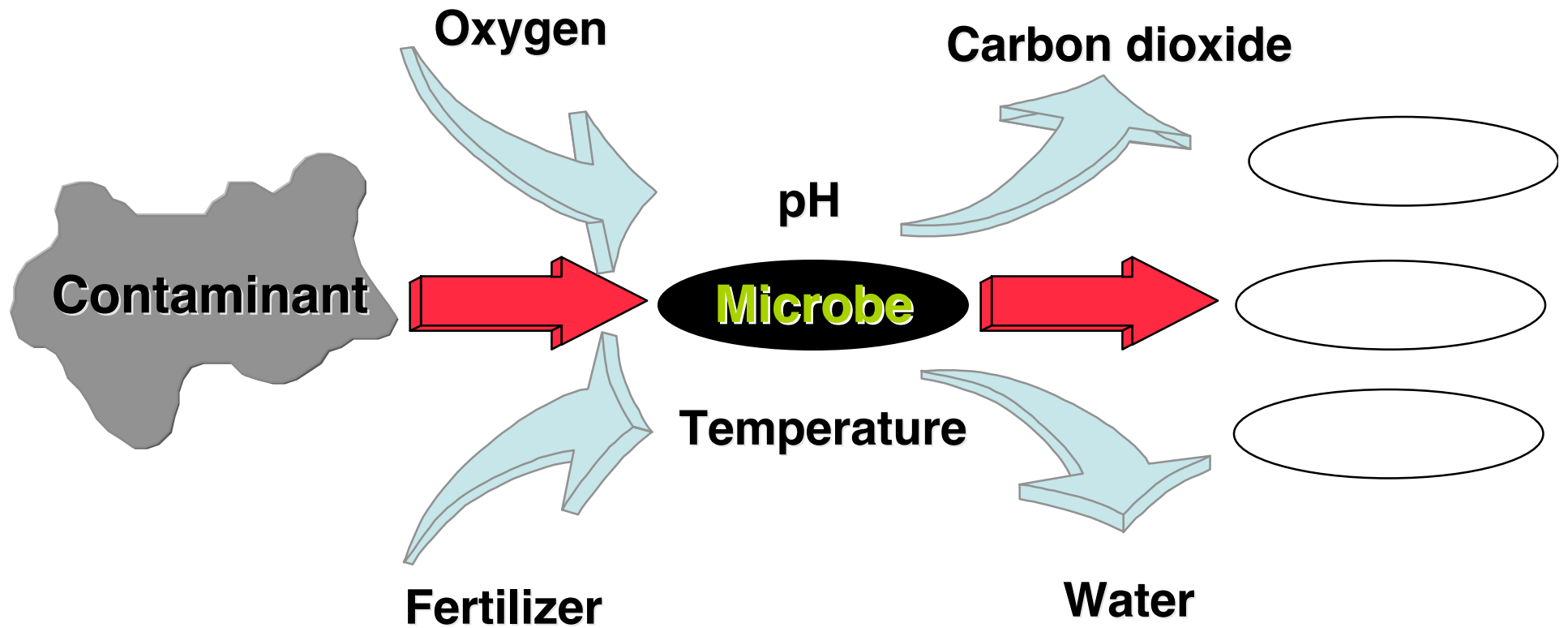
- molecular size, shape, charge and functional groups, concentration
- solubility in water; lipid/water partition coefficient
- solid/liquid/gas; volatilization
- toxicity
- possibility of spontaneous nonenzymatic reactions

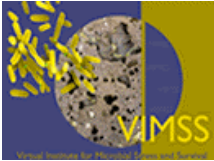
The Environment

- mechanical accessibility
- pH, pO_2 , temperature, redox potential
- presence of interfaces
- ionic composition and concentration
- water and wind speed, light quality and intensity
- presence of co-metabolites, essential nutrients, reactive radicals, etc.
- presence of appropriate organisms or plasmids



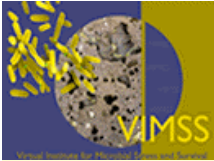
Bioremediation explained



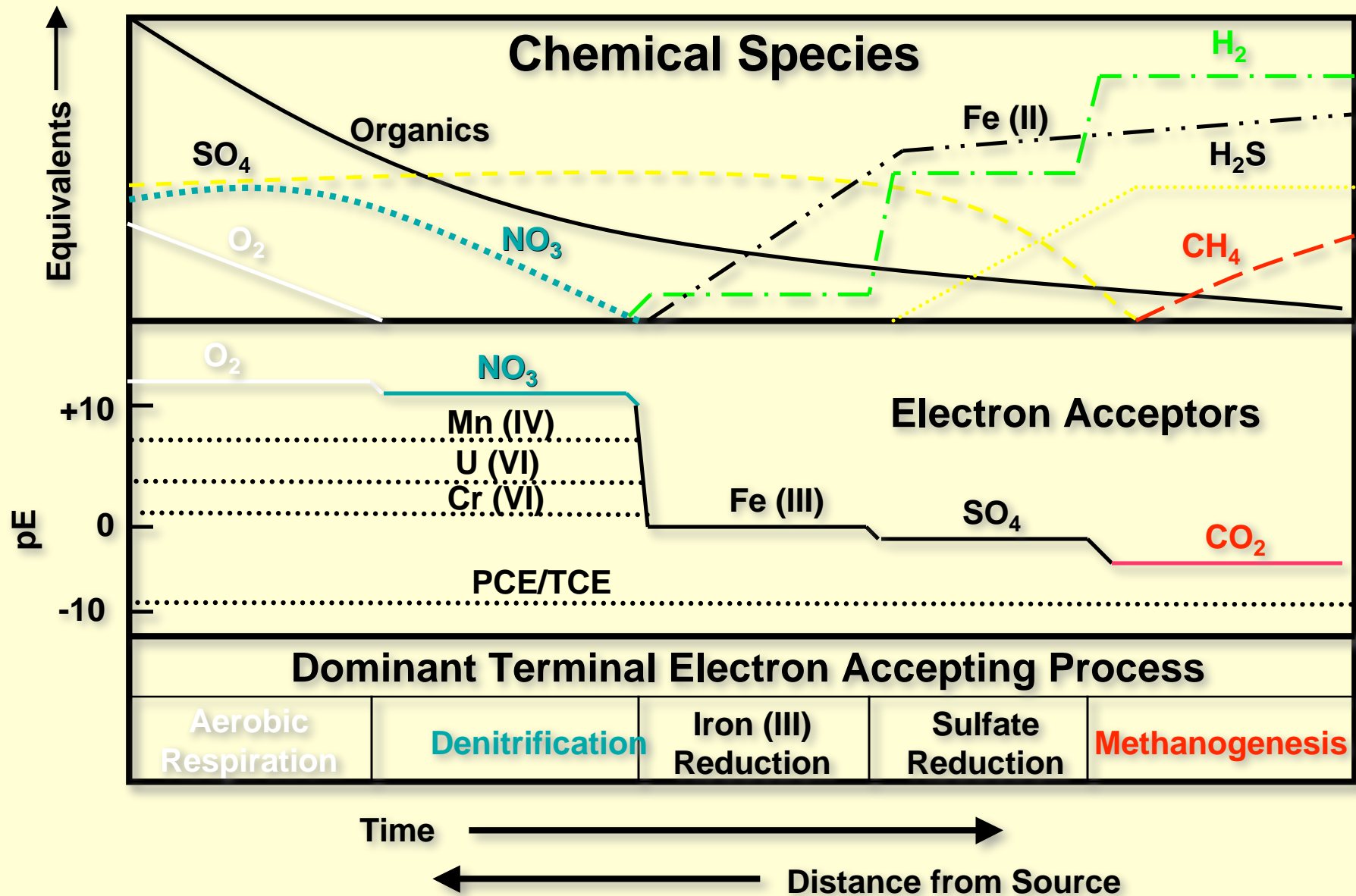


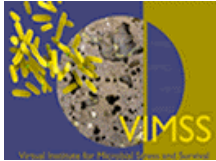
TEA and ED?

- Terminal Electron Acceptors (TEA)
 - Oxidizing agent in cellular respiration
 - O_2 , NO_3 , $Fe(III)$, SO_4 , CO_2
- Electron Donors (ED)
 - Reducing agents
 - Energy sources: usually carbon sources also e.g. sugars, etc.



Critical Biogeochemistry





Bioremediation Historical Perspective

prehistoric Fermentation (Second Oldest Profession?)

6000 BC Kitchen middens, compost piles

1900 BC Greeks walled refuse bioreactors

1891 First Waste Water Treatment Plant (Sussex, UK)

1946 Zobell Demonstrates Oil Biodegradation

1950 Petroleum Land-Farming Widely Used

1968 Bilge Water of Queen Mary Biotreated (Bioaugmentation)

1974 Raymond Patent for In Situ Biotreatment of Gas Spills

1981 First U.S. Patent on life (petroleum degrader) GE

1988 French Limited Superfund Site Test

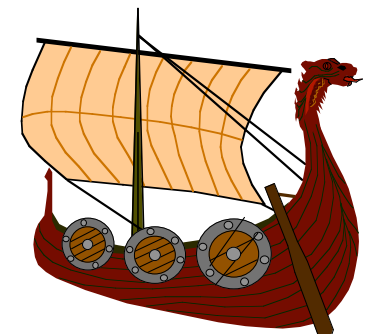
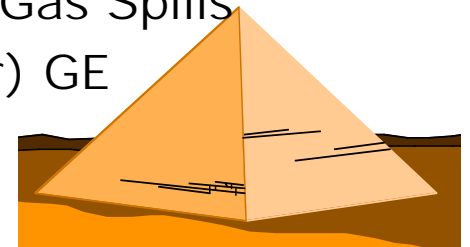
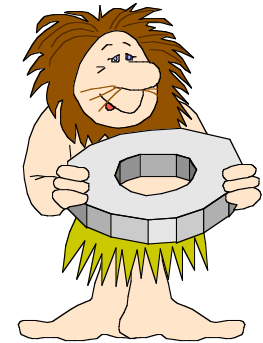
1989 Exxon Valdez Spill Demonstration by EPA

1992 SRS Integrated Demonstration for TCE/PCE

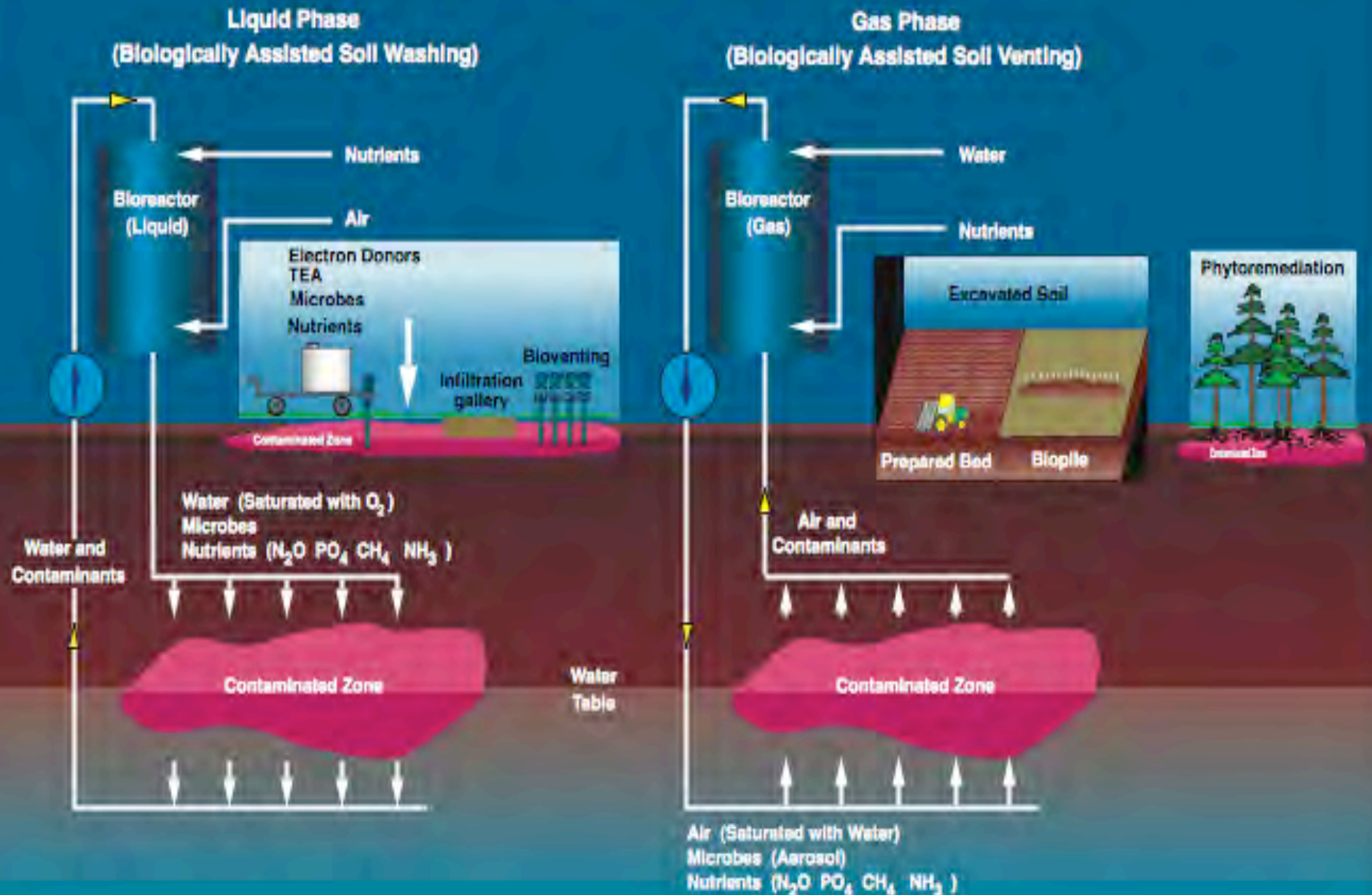
1993 GE Hudson River Caisson Demonstration for PCB

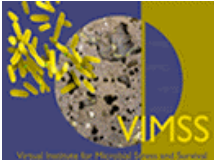
1997 UT/ORNL lysimeter tests of GMO

1999 Oyster Site release of Adhesion-less strain



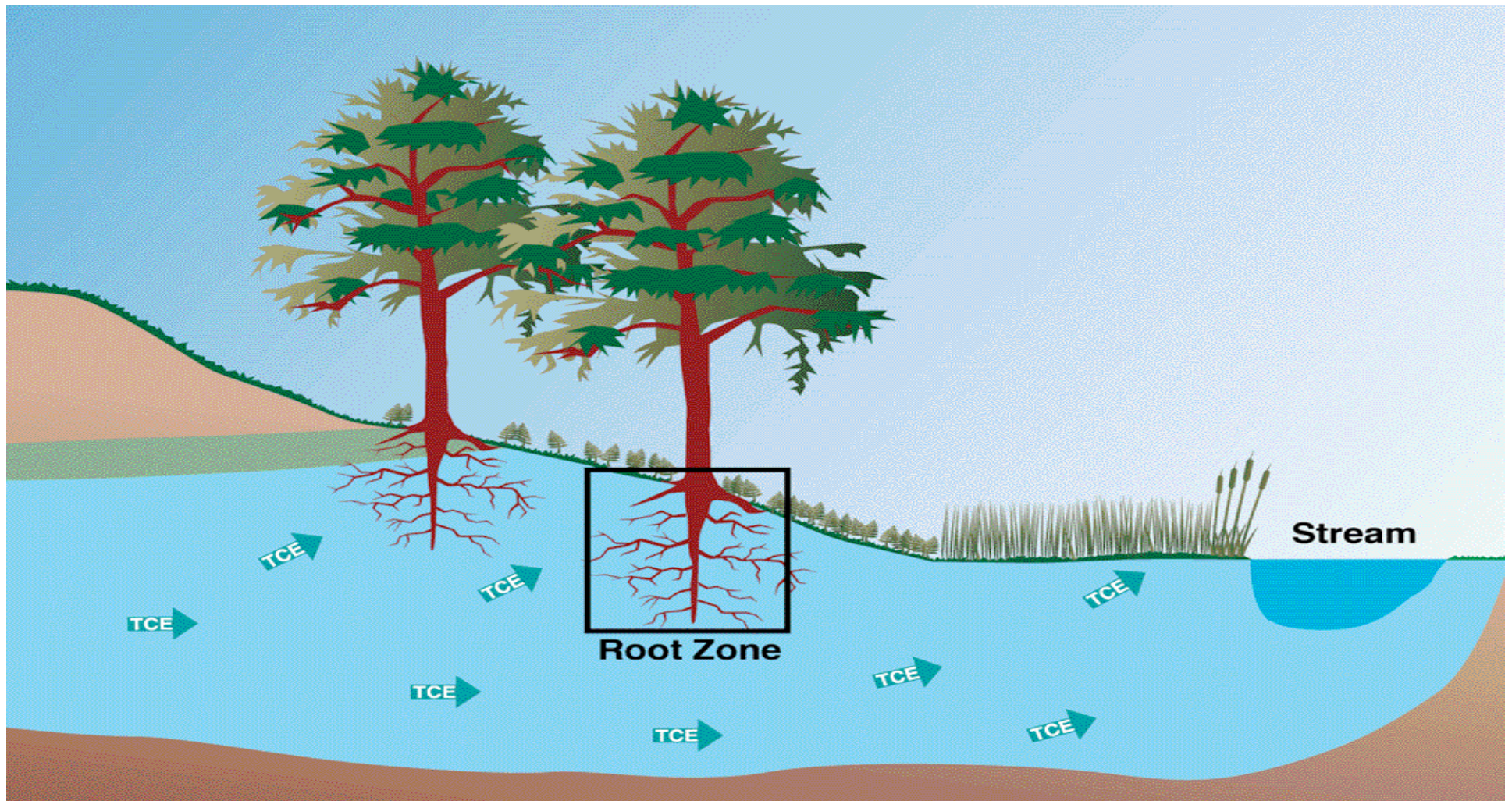
Bioremediation Technologies

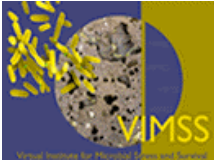




Intrinsic Bioremediation

Unmanipulated, unstimulated, unenhanced biological remediation of an environment; i.e. biological *natural attenuation* of contaminants in the environment.





Land Farming

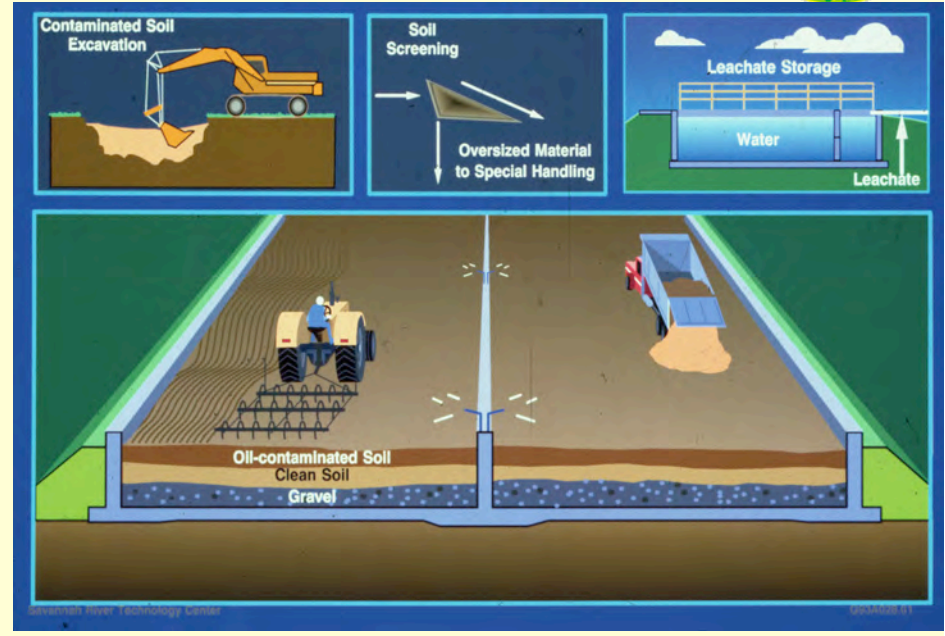
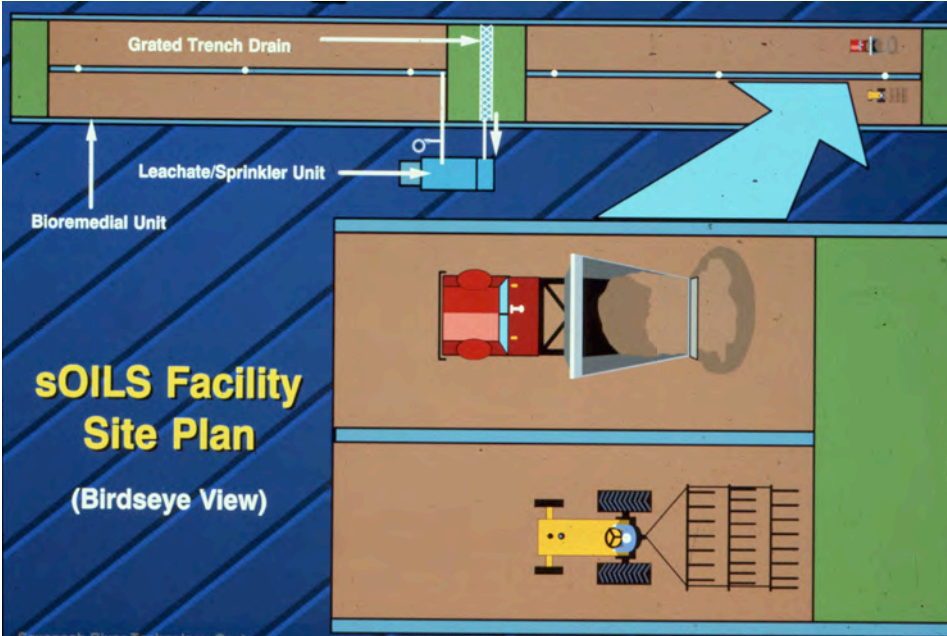
Biologically treating uncontained surface soil, usually by aeration of the soil (tilling) and addition of fertilizer or organisms, hence farming.

	Prepared Bed	Bioventing
Various (Bartha, 1986)	52–641	
SRS	10–107	10–65
Poland (refinery biopile)	33–121	
Italy (biopile – crude)	60	
Hill AFB, Utah		10
Tyndall AFB, Florida		2–20
The Netherlands		2–5
The Netherlands		8
Patuxent River NAS, Maryland		3
Fallon NAS, Nevada		5
Eicklson AFB, Alaska		1–10
Kenai, Alaska		21
Tinker AFB, Oklahoma		2.7–18

*all values in mg TPH/kg soil/day



sOILS Facility





Polish Refinery

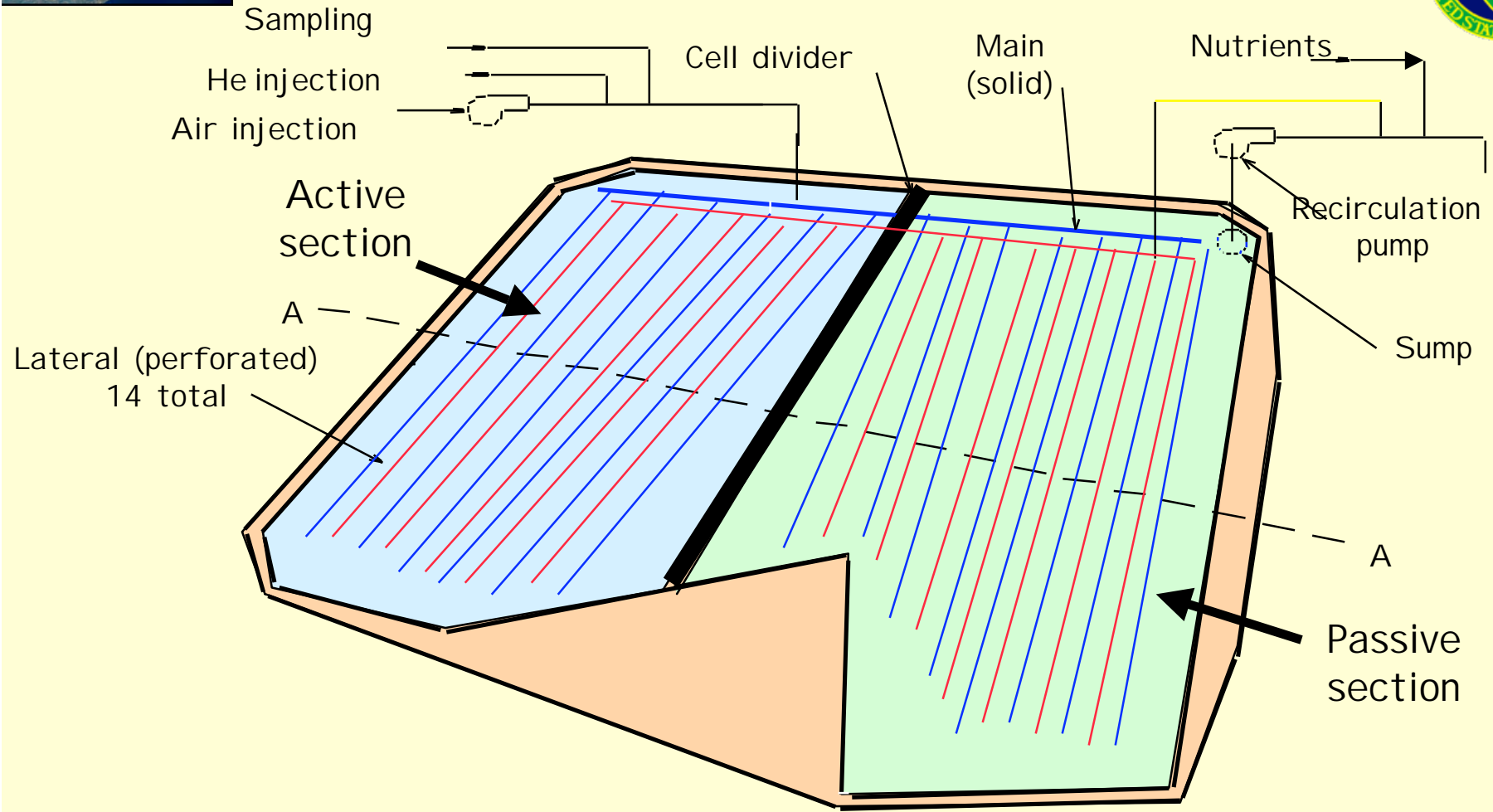


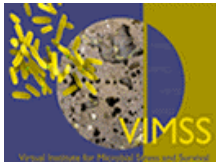
Before
4167 yd³ of sludge
contaminated soil

After

18 Months
120 metric tons
destroyed (81%)
Green Zone





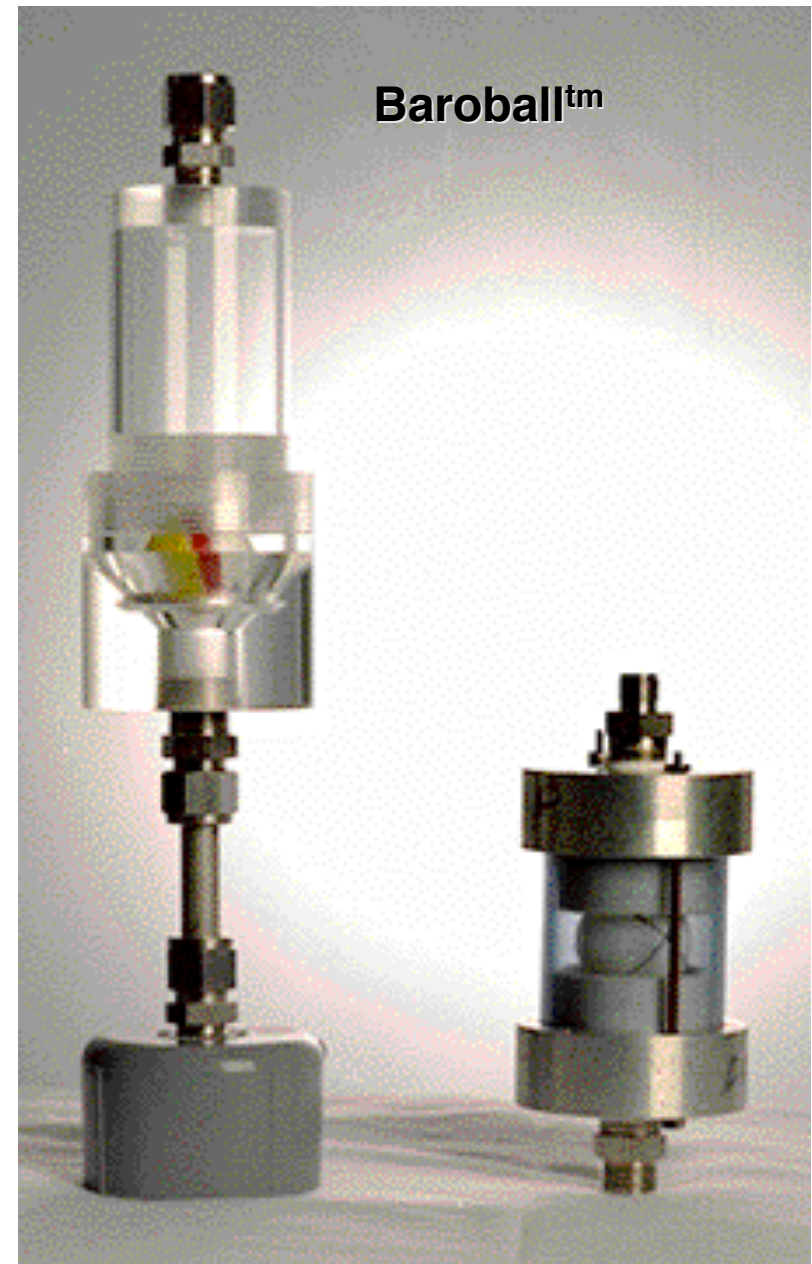


Passive Bioremediation

Using natural processes for biostimulation, e.g. barometric pumping, natural infiltration, to deliver nutrients or manipulate the environment, i.e. engineering controls

Campaign	Passive	Active
OC-1	44*	119
OC-2	82	94
OC-3	33	0
OC-4	0	37
OC-5	60	121

*mg TPH/kg Soil/day



Model Assumptions

$$m(t) = M/R^3(R^2 - 2a\Delta ct/\gamma)^{3/2}$$

NAPL (fraction A) content	~ 40% of total TPH inventory in soil
Readily available fraction content	~ 45% of total TPH inventory in soil
Sorbed fraction content	~ 15% of total TPH inventory in soil
Soil porosity:	= ~ 0.3

Characteristics of NAPL fraction (**Fraction A**):

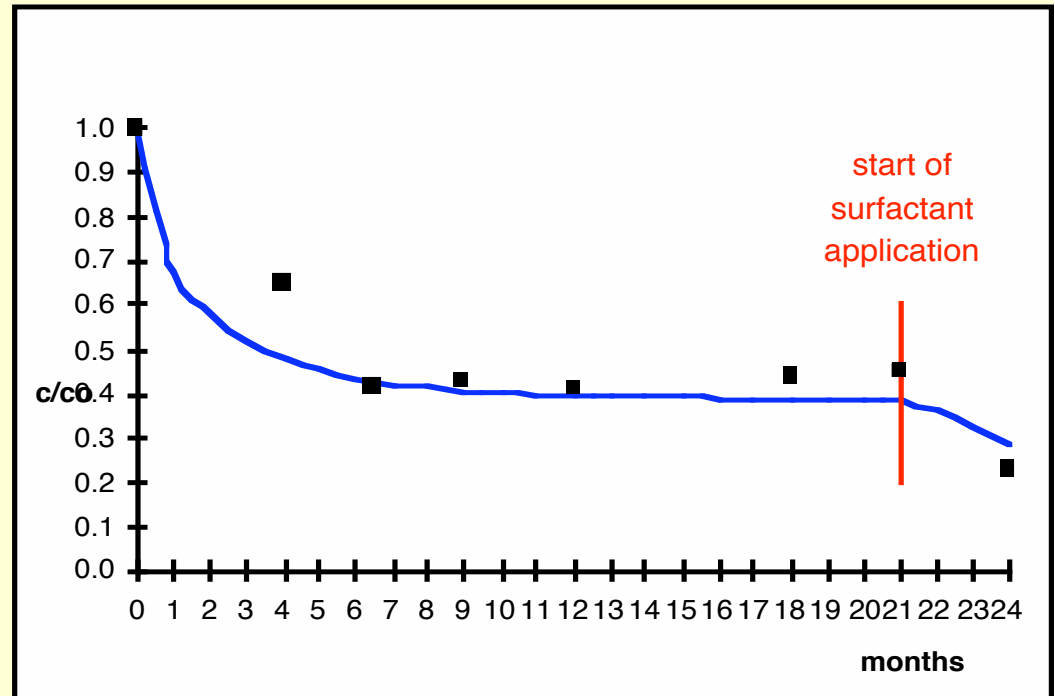
Average radius of aggregates (droplets)	R = 1.0 cm
Solubility in water	c = 10 mg/l before the surfactant was added c = 100 mg/l after the surfactant was added

Characteristics of readily available fraction (**Fraction B**):

Average radius of soil aggregates:	$r_0 = 1.0$ cm
Desorption coefficient	$K_d = 100$
Pore diffusivity of contaminant	$D_{eff} = 5 \times 10^{-11}$ cm ² /s
Liquid mass transfer coefficient	$k_l = 1 \times 10^{-5}$ cm/s

Characteristics of sorbed fraction (**Fraction C**):

Average radius of soil aggregates:	$r_0 = 30$ m
Desorption coefficient	$K_d = 1 \times 10^5$
Pore diffusivity of contaminant	$D_{eff} = 5 \times 10^{-13}$ cm ² /s
Liquid mass transfer coefficient	$k_l = 1 \times 10^{-5}$ cm/s



Biostimulation - The addition of organic or inorganic compounds to cause indigenous organisms to effect remediation of the environment, e.g. fertilizer, surfactants.

Bioaugmentation - The addition of organisms to effect remediation of the environment, e.g. contaminant-degrading bacteria injection into an aquifer, GMO.



Biostimulation Requirements



1. correct microbes must be present
2. ability to stimulate target microbes
3. ability to deliver nutrients
4. C:N:P - 30:5:1 for balanced growth (Paul and Clark, 1989) 100:10:2 in field practice (Litchfield, 1993)

Gases: air, oxygen, nitrous oxide, propane, methane, triethyl phosphate, etc.

Liquids: lactic acid, molasses, vegetable oil, acetate, Chitin, hydrogen release compound (HRC[®]), MRC[®], etc.

Solids: bulking agents (saw dust, agricultural byproducts), oxygen release compound (ORC[®]), etc.





D Area Oil Seepage Basin

2 trenched horizontal wells at 3 m

1 blower (200 scfm)

Methane, N₂O, TEP

In less than 6 months



Methylene Chloride: 2300 ppb to < 2 ppb

Vinyl Chloride: 300 ppb to < 5 ppb

Dichloroethylene: 100 ppb to < 2 ppb

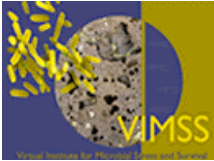
Trichloroethylene: 100 ppb to < 5 ppb

Tetrachloroethylene: 50 ppb to < 10 ppb

BTEX: 50 ppm to < 1 ppm

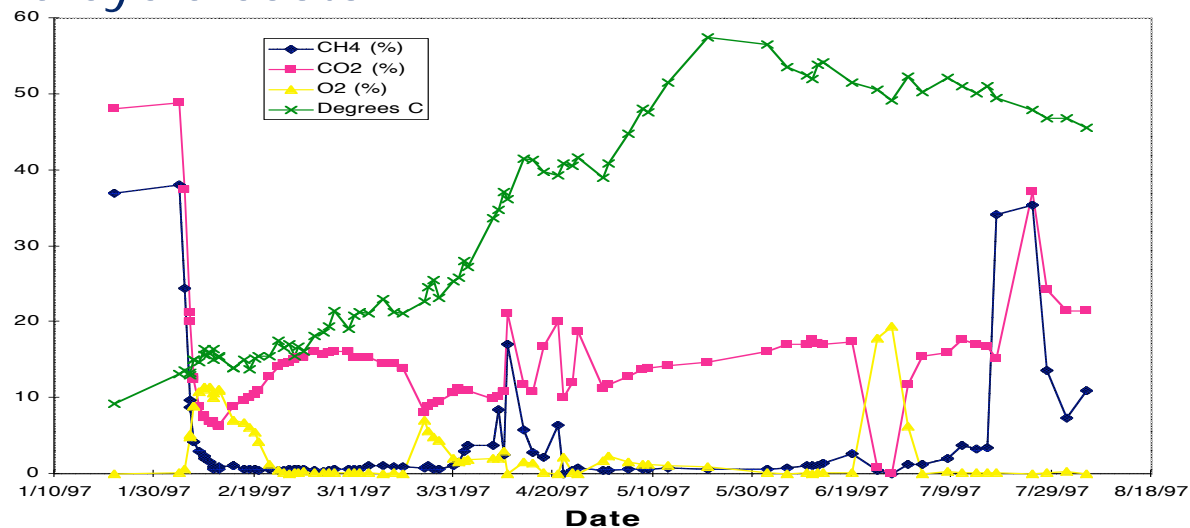
No Action ROD filed 6/98, granted 3/99

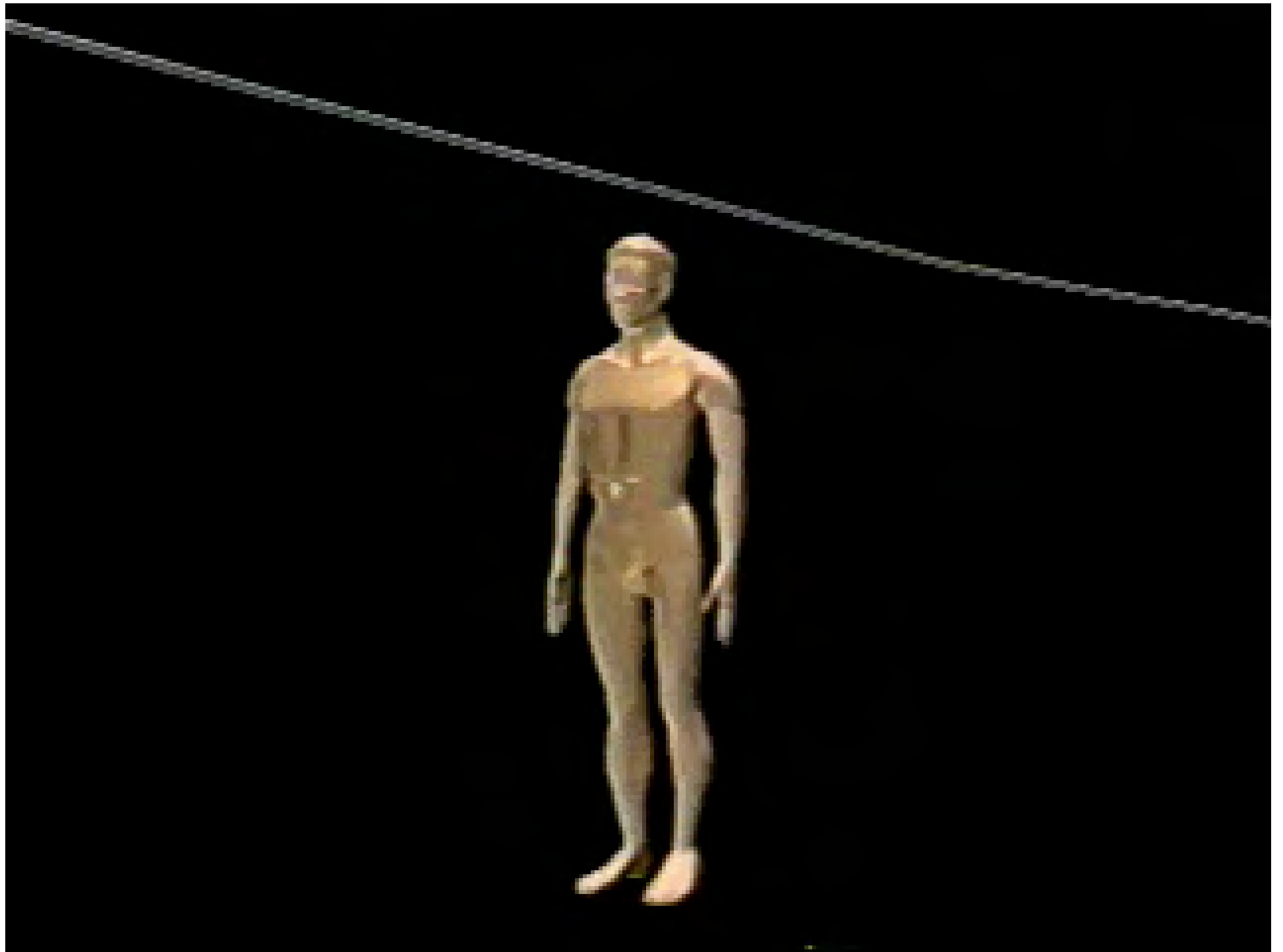




Aerobic Landfill Bioremediation

- increased biodegradation rate
- increased subsidence
- eliminated need for leachate treatment
- stabilized refuse mass sooner
- decreased long-term liability and monitoring costs
- decreased leaching of metals and organic contaminants
- decreased methane generation
- Reduced life cycle costs





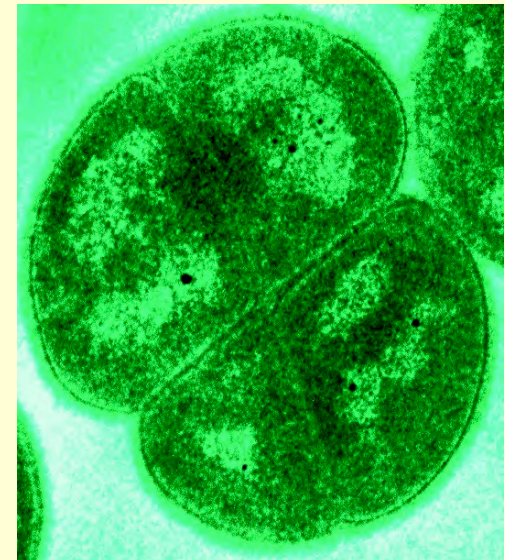
Bioaugmentation Advantages

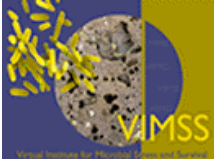


1. "new" spills where microflora has not had time to adapt or grow (vector)
2. recalcitrant contaminants (GMO)
3. biomass can not establish or maintain itself (GMO)
4. biobarrier (ultramicrobacteria, GMO)
5. controlled environment (GMO)

Pseudomonads (oil spills) - several commercial products

Dehalococcoides ethenogenes
(chlorinated solvents) new products
from Regenesis and GeoSyntec





Systems Biology Approach

Ecosystem

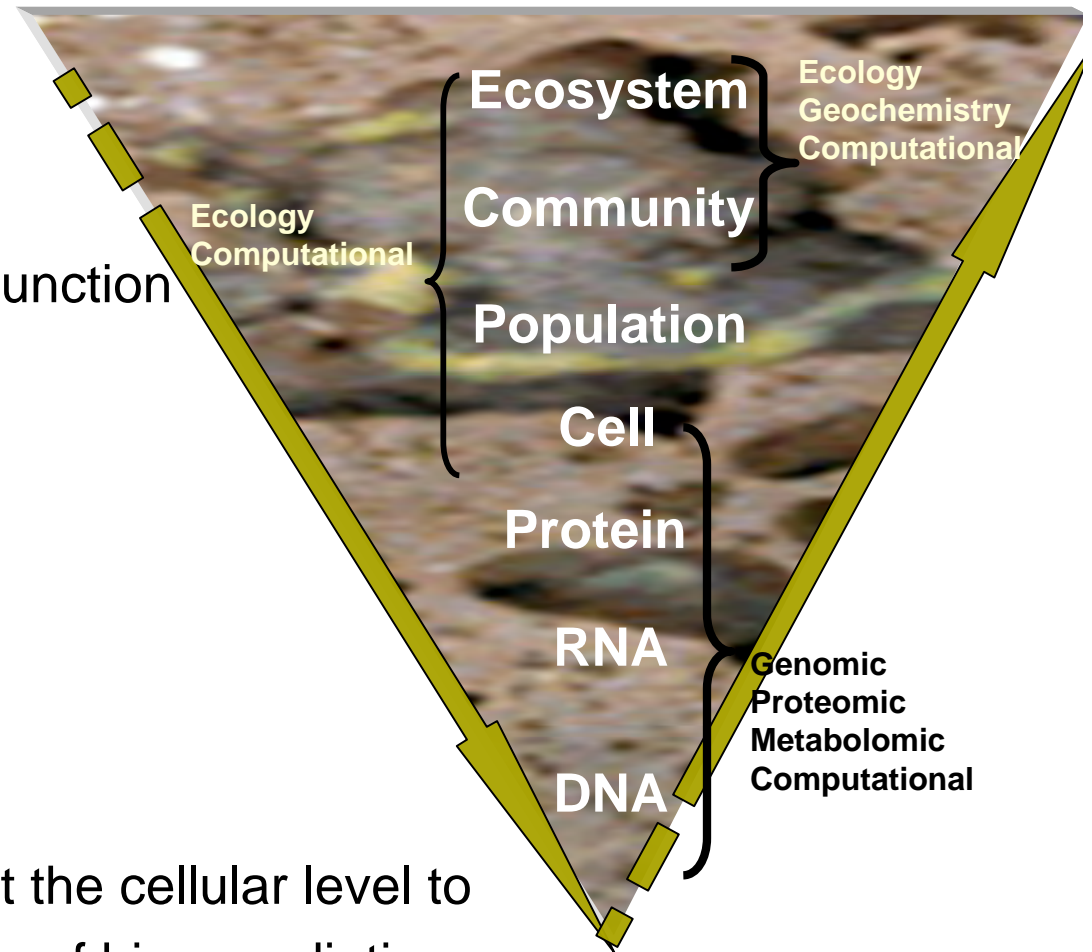
Identify key factors (i.e., stresses) that drive community structure and composition and impact the survival and efficacy of heavy metal-reducers

Community and Population

Understand impacts on structure/function relationships

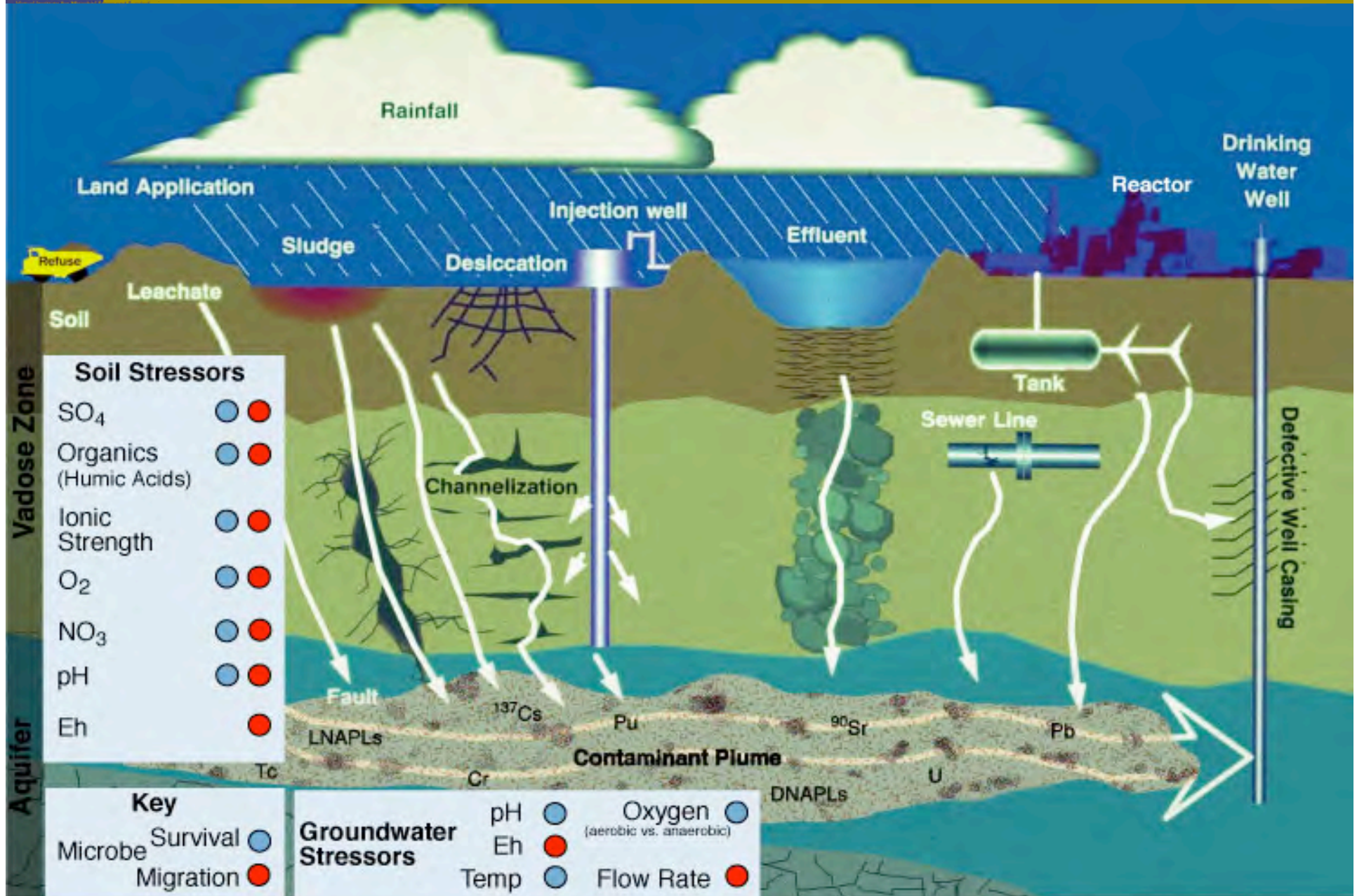
Cell

Analyze DNA, RNA, and protein at the cellular level to understand cellular effects in terms of bioremediation

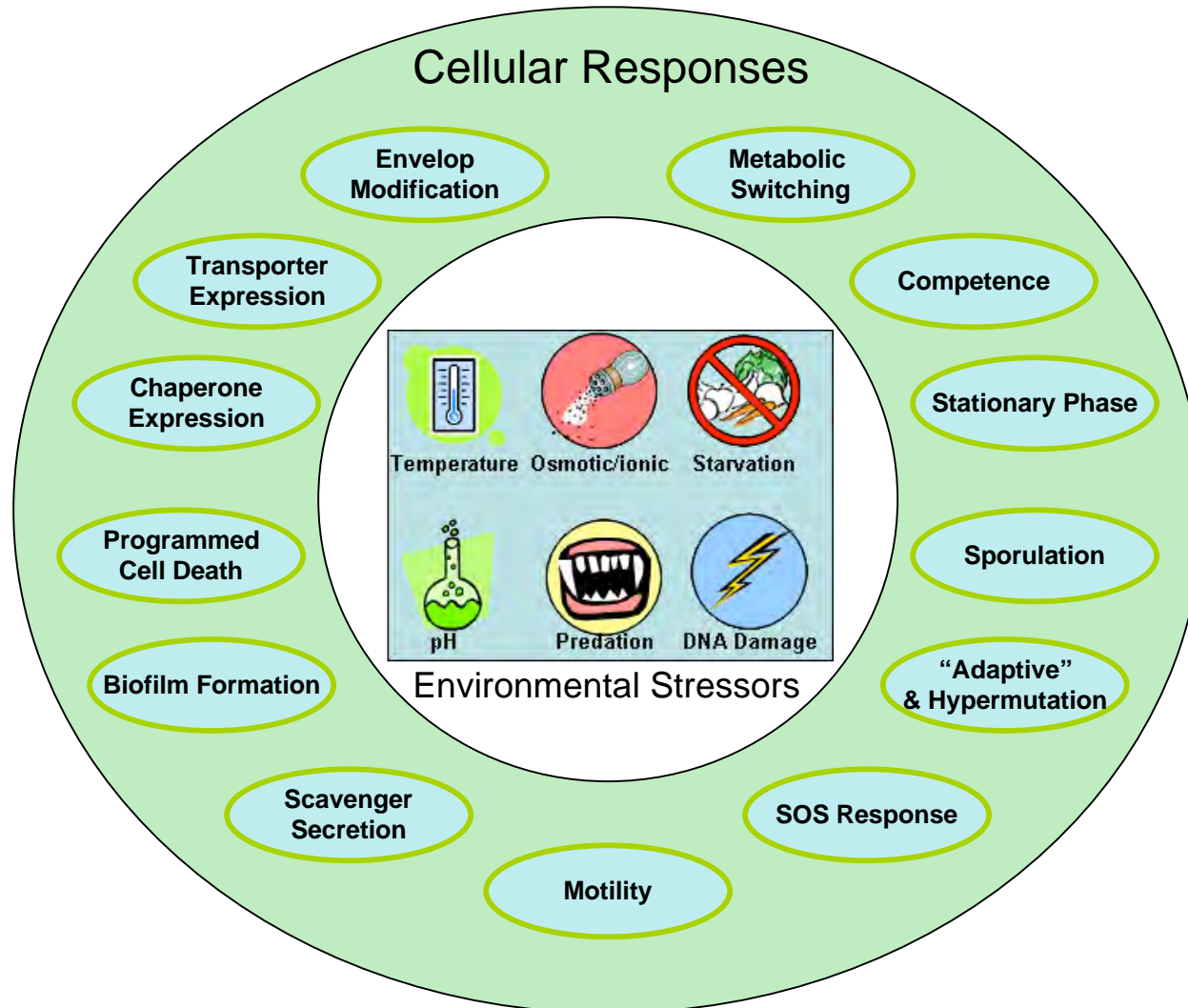




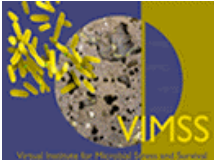
Stress Response Pathways in Biogeochemistry



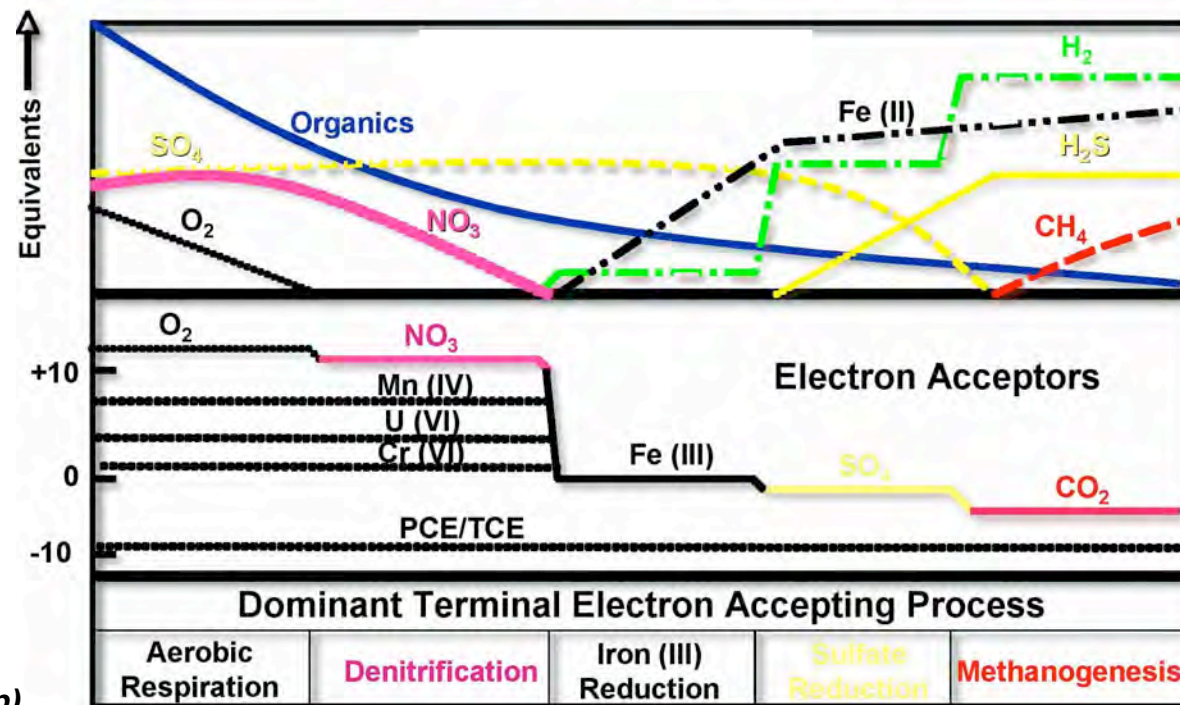
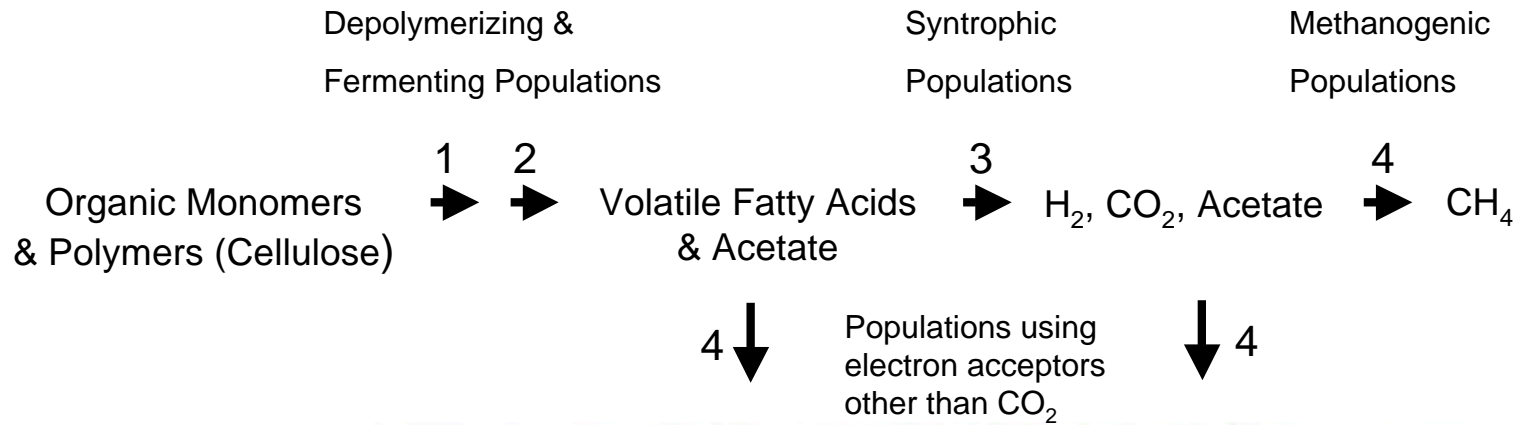
Relevant Stress Responses



(Hazen & Stahl, 2006 - Current Opinions in Biotechnology)



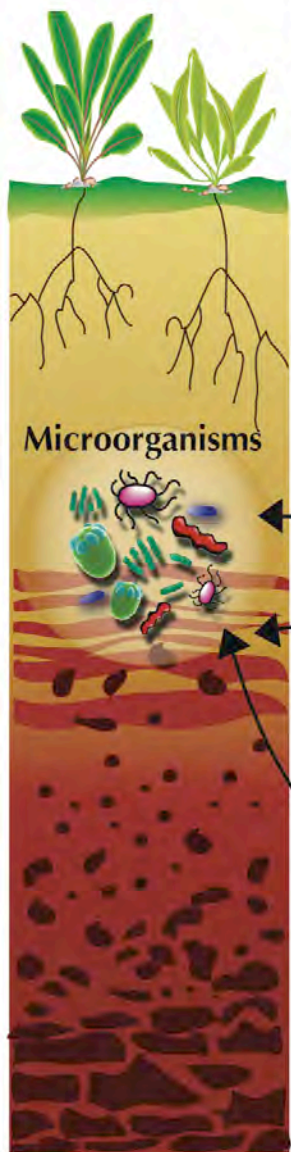
Multiple Trophic Interactions Determine Delivery of Electron Donors to Terminal Electron Accepting Species



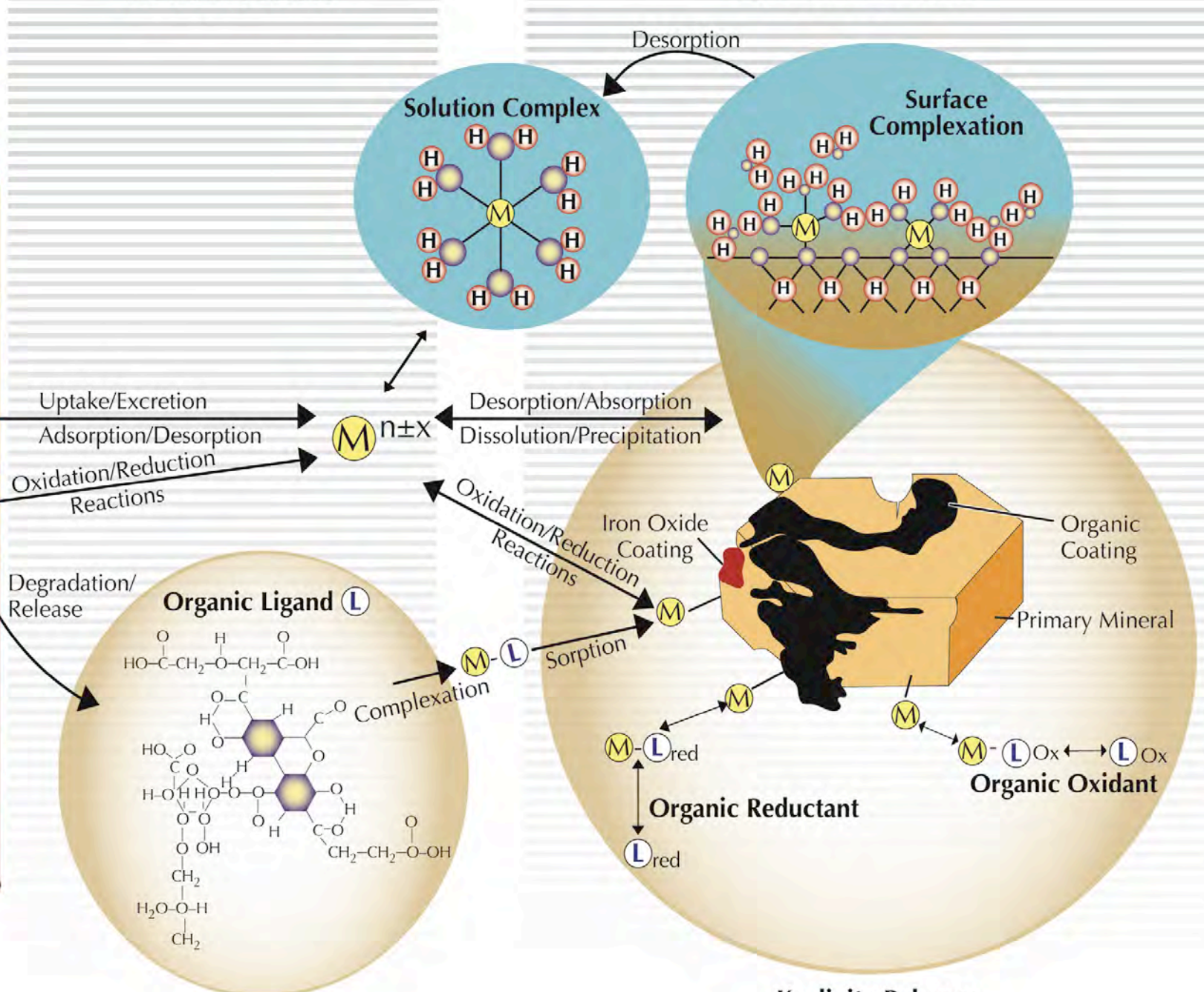
(Hazen and Stahl, in prep)

Biotic Processes

Abiotic Processes



Microorganisms



Solution Complex

Surface Complexation

Desorption

Uptake/Excretion

Adsorption/Desorption

Oxidation/Reduction
Reactions

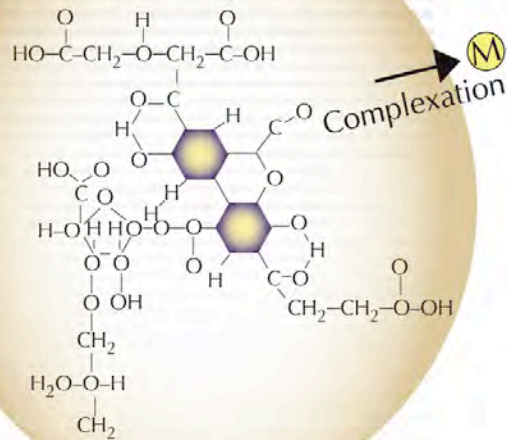
Desorption/Absorption

Dissolution/Precipitation

Oxidation/Reduction
Reactions

Degradation/
Release

Organic Ligand (L)



Complexation

Sorption

Iron Oxide
Coating

Organic
Coating

Primary Mineral

$M-L_{red}$

L_{red}

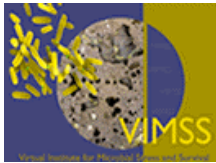
Organic Reductant

$M-L_{Ox}$

L_{Ox}

Organic Oxidant

Kaolinite-Polymer
Complexes



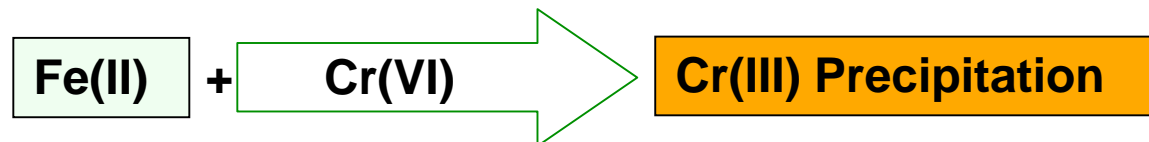
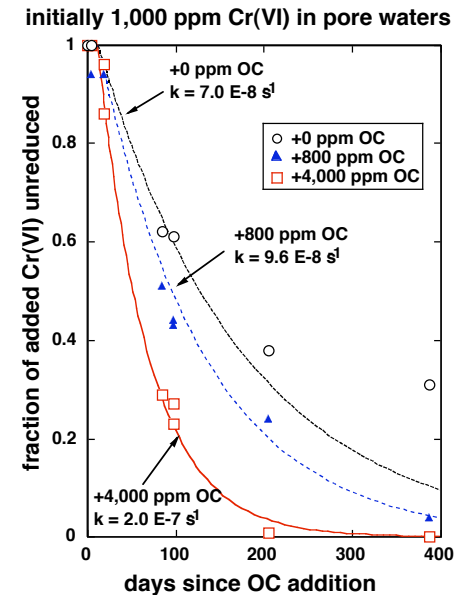
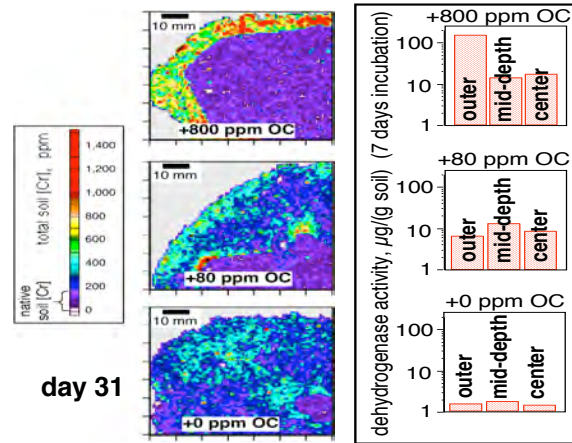
Mesoscale Studies on Cr(VI) Bioreduction Lab Studies

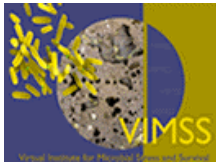
Jiamin Wan, Tetsu Tokunaga, Mary Firestone, Eoin Brodie and Terry Hazen (ERSP/NABIR supported 1998-2004)

Tokunaga, T. K. J. Wan, M. K. Firestone, T. C. Hazen, K. R. Olson, D. J. Herman, S. R. Sutton, and A. Lanzirotti. 2003. *In-situ* reduction of Cr(VI) in heavily contaminated soils through organic carbon amendment. *J. Environ. Qual.* 32:1641-1649.

Tokunaga, T. K., J. Wan, T. C. Hazen, E. Schwartz, M. K. Firestone, S. R. Sutton, M. Newville, K. R. Olson, A. Lanzirotti, and W. Rao. 2003. Distribution of chromium contamination and microbial activity in soil aggregates. *J. Environ. Qual.* 32:541-549.

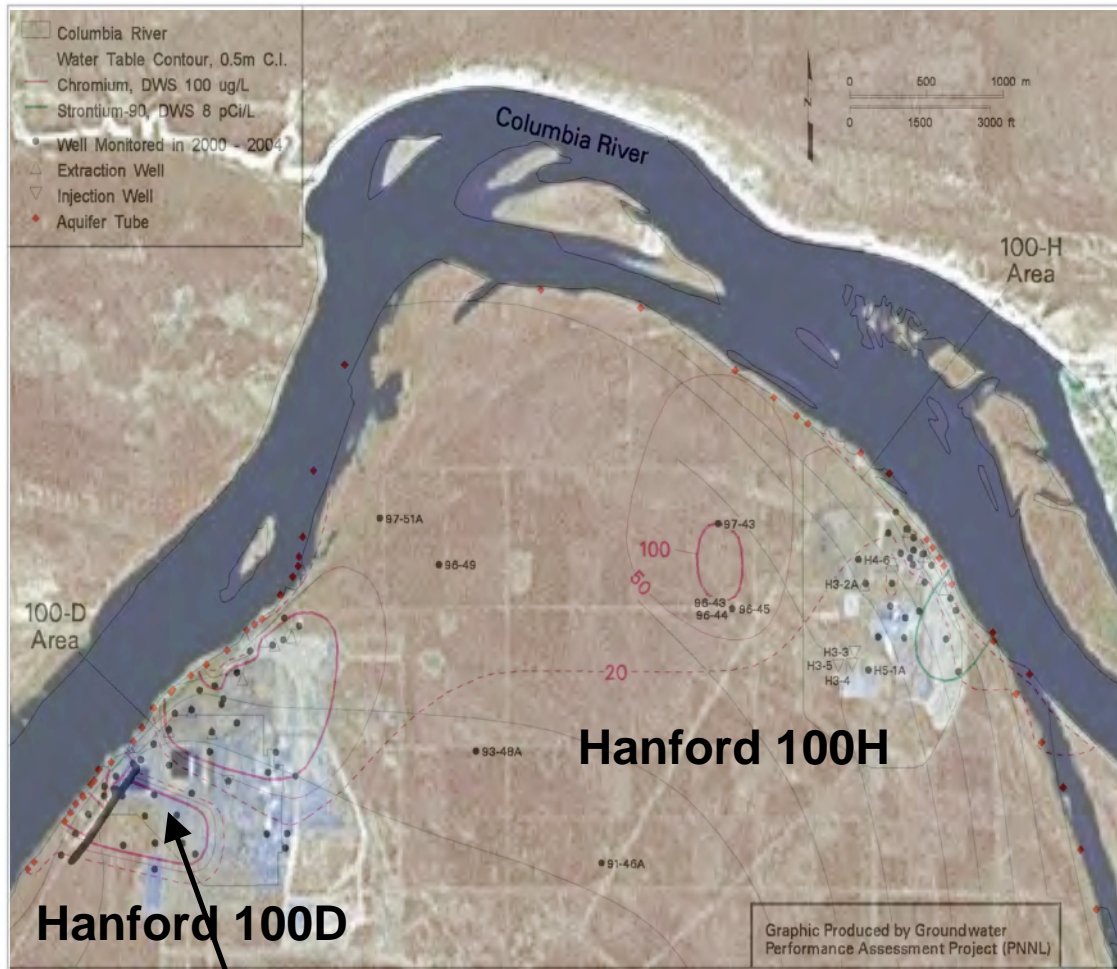
Tokunaga, T. K., J. Wan, M. K. Firestone, T. C. Hazen, E. Schwartz, S. R. Sutton, and M. Newville. 2001. Chromium diffusion and reduction in soil aggregates. *Environmental Science & Technology* 35:3169-3174.





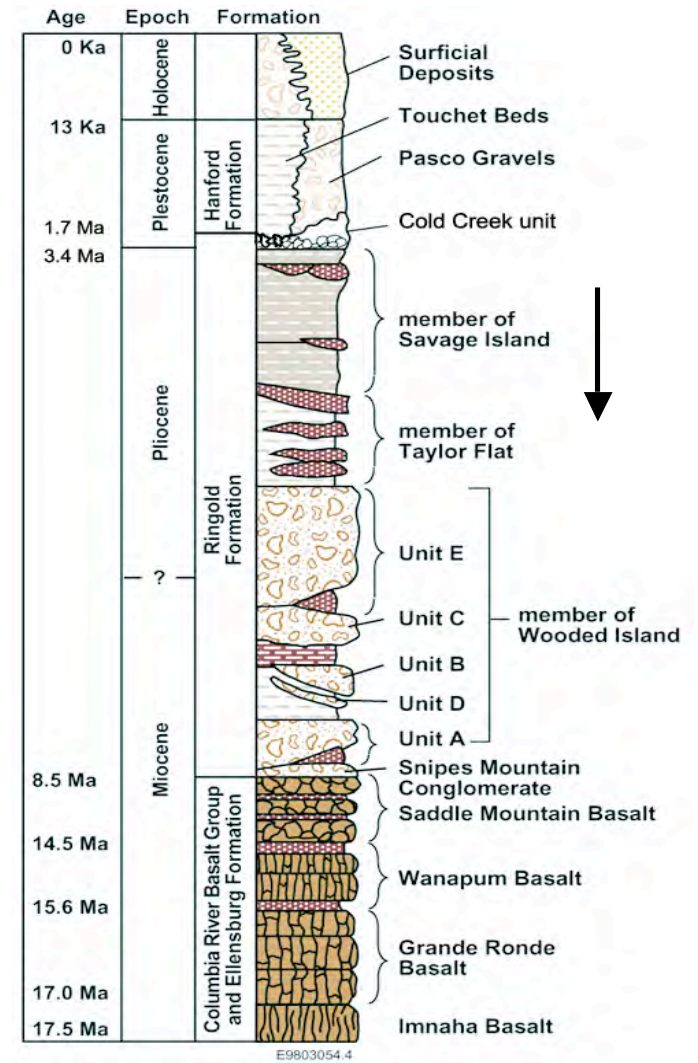
Hanford 100H Site Characterization

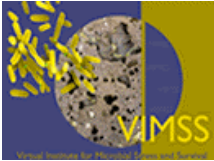
Cr Concentration Map



The Cr source is believed to be sodium dichromate ($\text{Na}_2\text{Cr}_2\text{O}_7 \cdot 2\text{H}_2\text{O}$)

Lithological Column

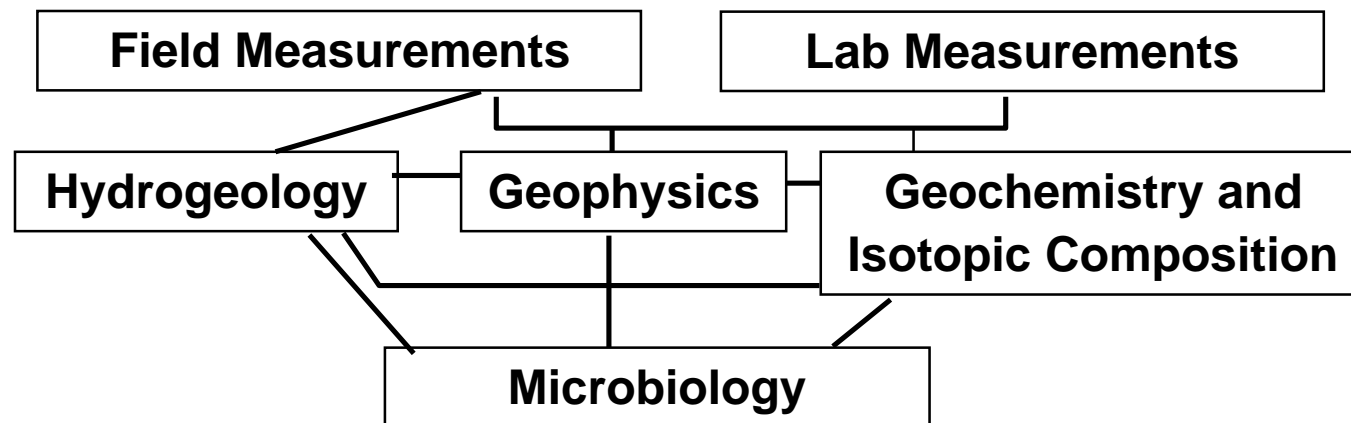


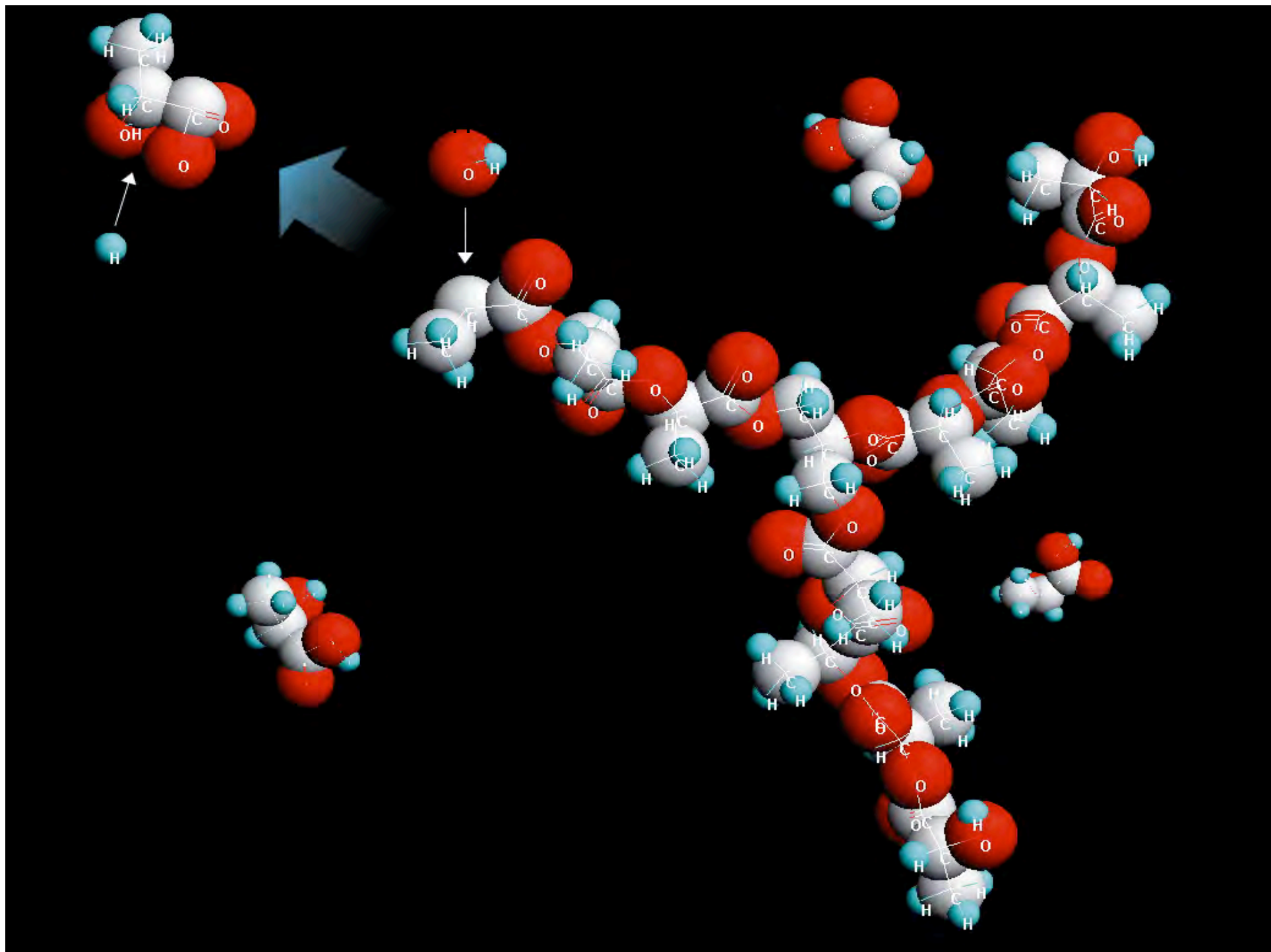


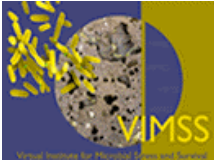
Overall Objective

To carry out field investigations to assess the potential for immobilizing Cr(VI) in groundwater using lactate-stimulated bioreduction of Cr(VI) to Cr(III) at the Hanford 100H site, and to determine critical community structure changes and stressors that would enable control and predictions of fundamental biogeochemistry that enables this bioremediation strategy for Cr(VI)

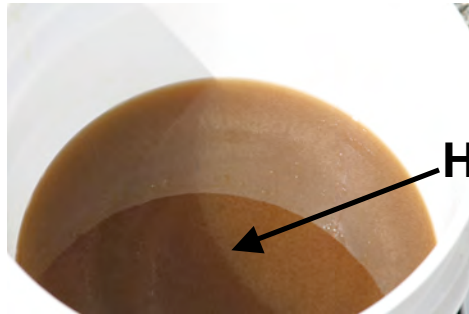
Integrated Approach







Field HRC Injection Test



Injection of 40 lbs of ^{13}C -labeled HRC
Well 699-96-45, August 3, 2004

Pumping - 27 days
Well 699-96-44

**Injection at depths
of 44 ft to 50 ft**

**Hanford sandy gravel
and gravelly sand**

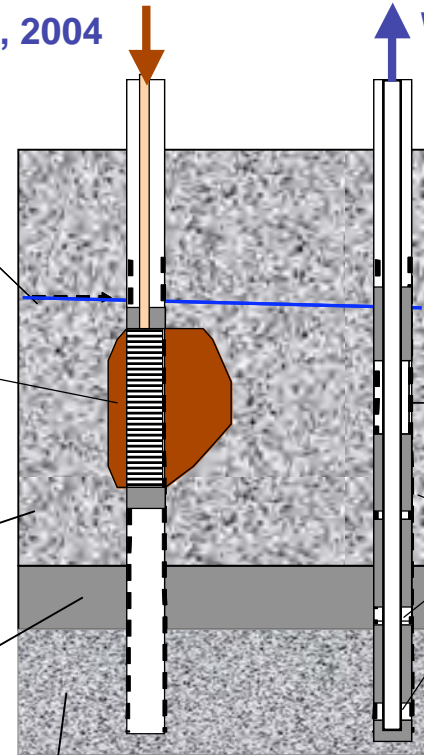
Ringold clay

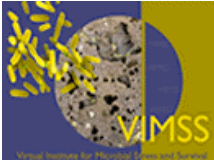
Ringold silt

Groundwater level

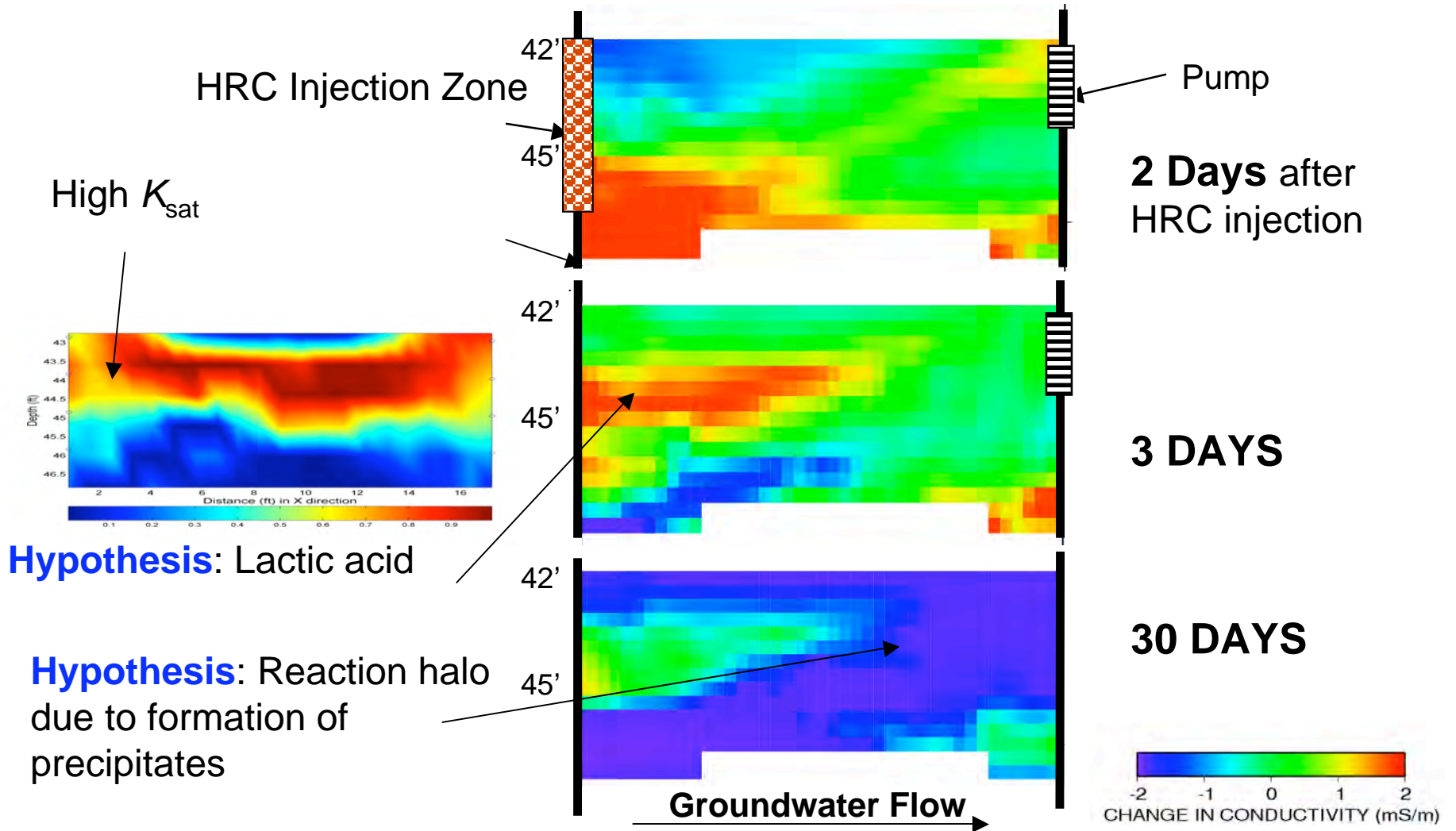
Pumping

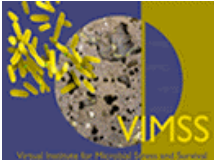
**Water
samplers**





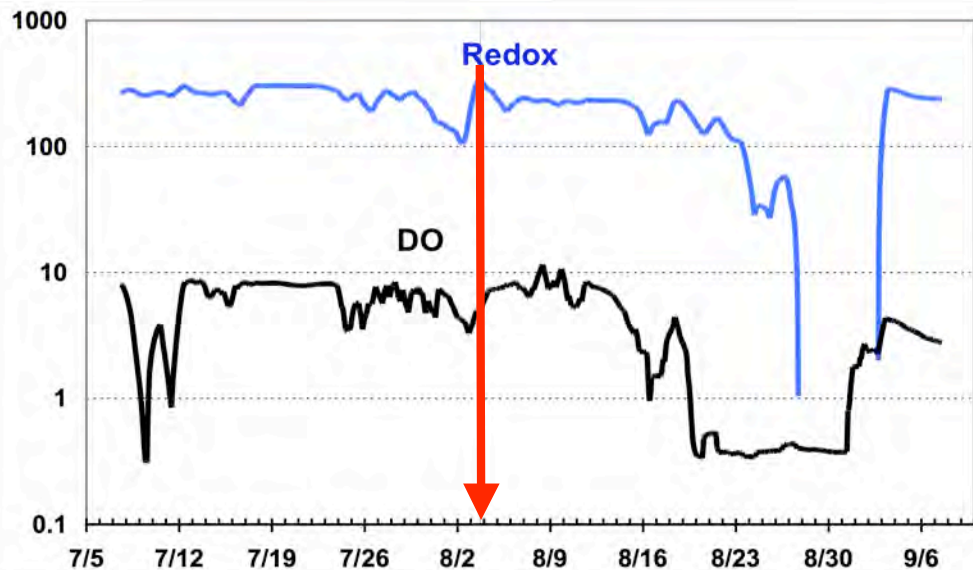
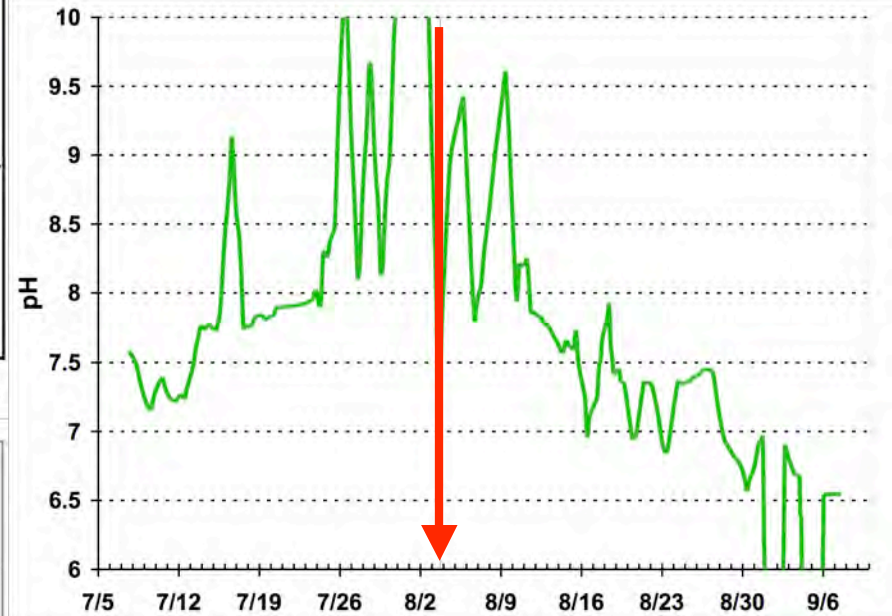
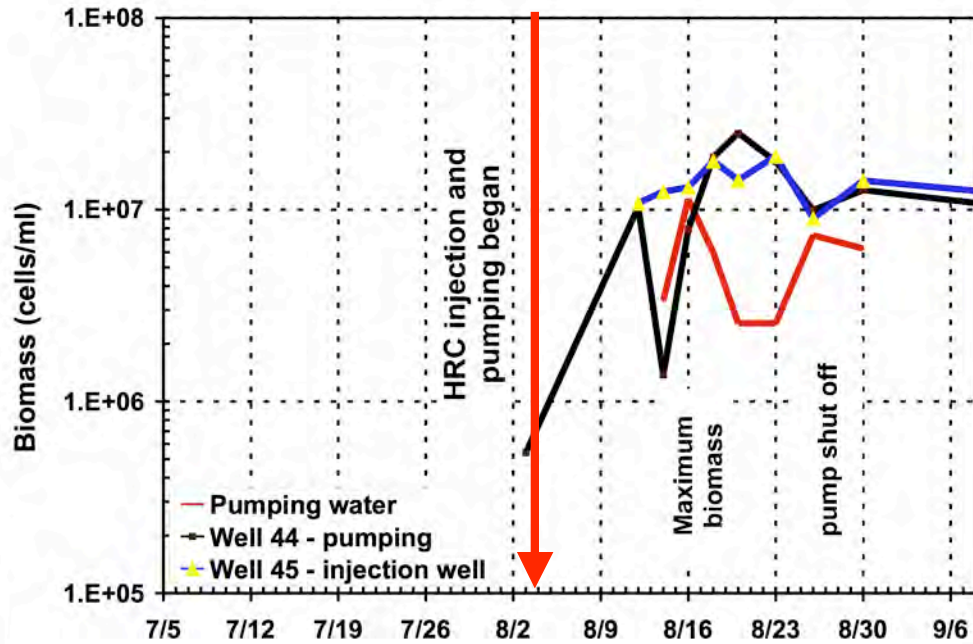
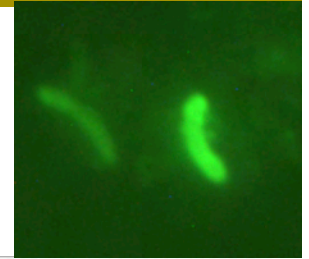
Post-HRC Injection Changes in Electrical Conductivity





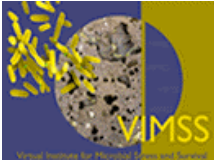
Results of HRC Biostimulation

D. vulgaris (direct fluorescent antibody)



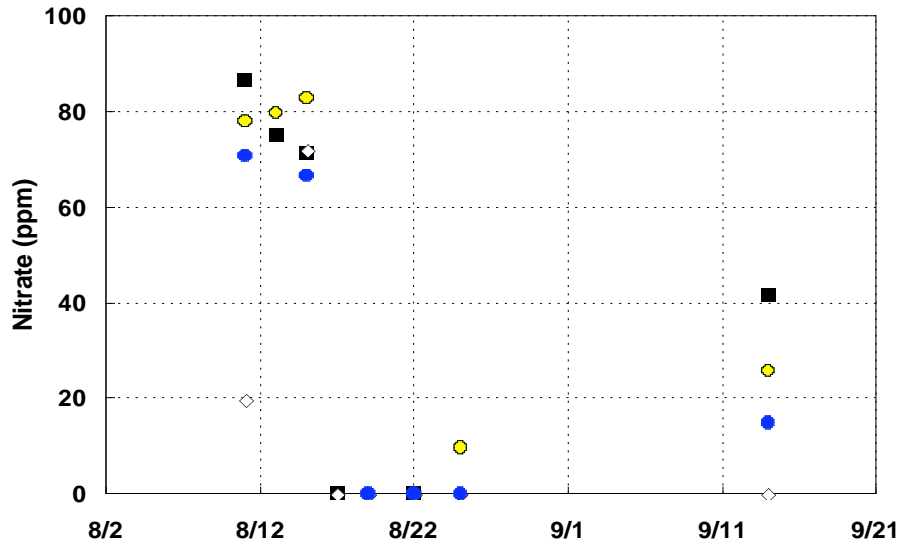
Redox dropped from 240 to -130 mV

DO dropped from 9 mg/l (~100%) to 0.35 mg/l (4.5%)

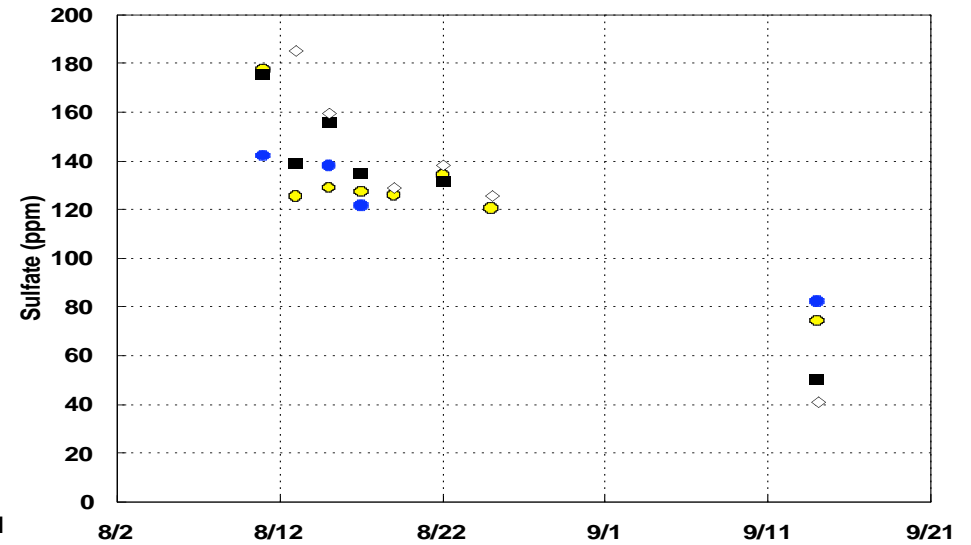


Biogeochemical Evidence of Microbial Metabolism in Groundwater

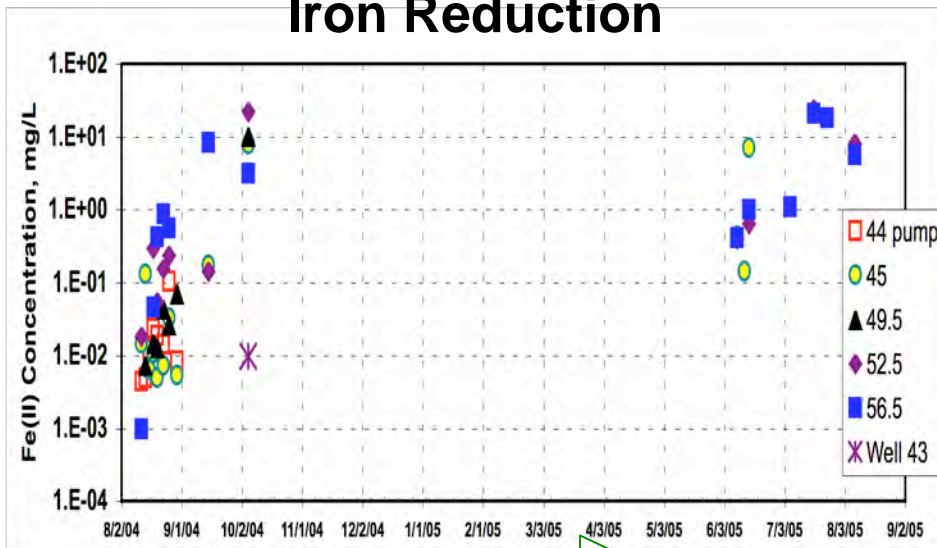
Denitrification



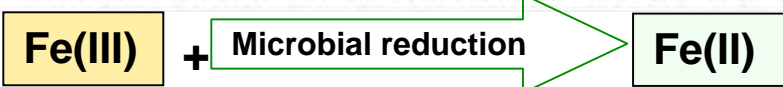
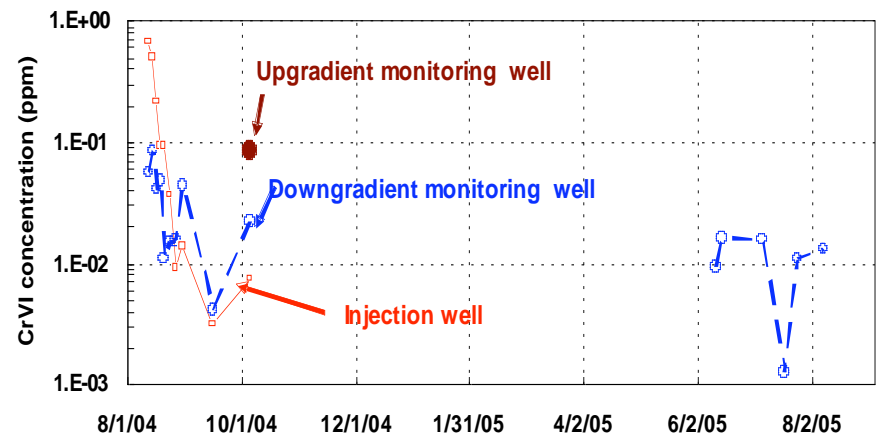
Sulfate reduction

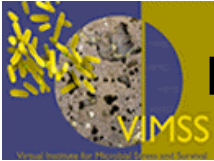


Iron Reduction

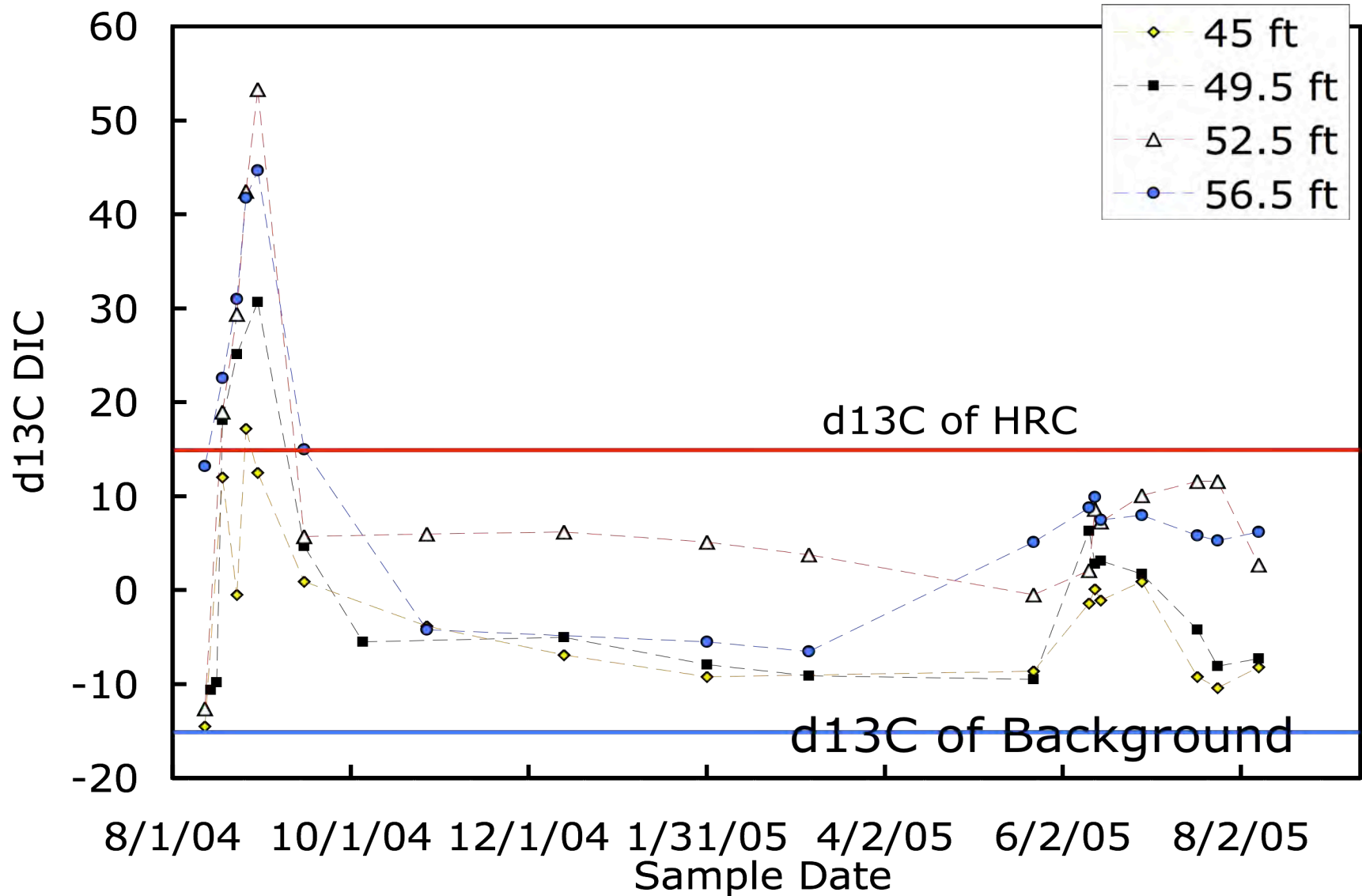


Average Soluble Cr(VI) Concentration

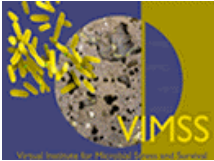




Biogeochemical Evidence of Microbial Metabolism in Groundwater

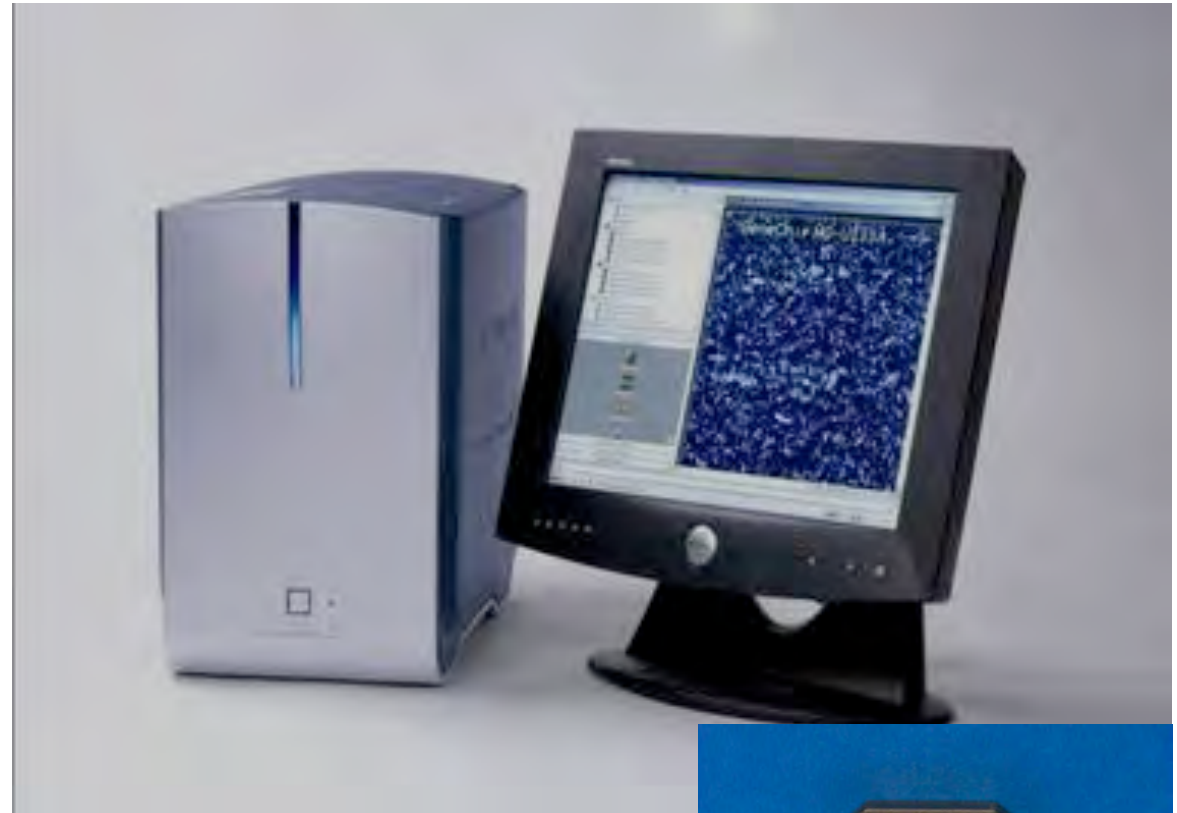


$\delta^{13}\text{C}$ of Dissolved Inorganic Carbon is Byproduct of HRC Metabolism



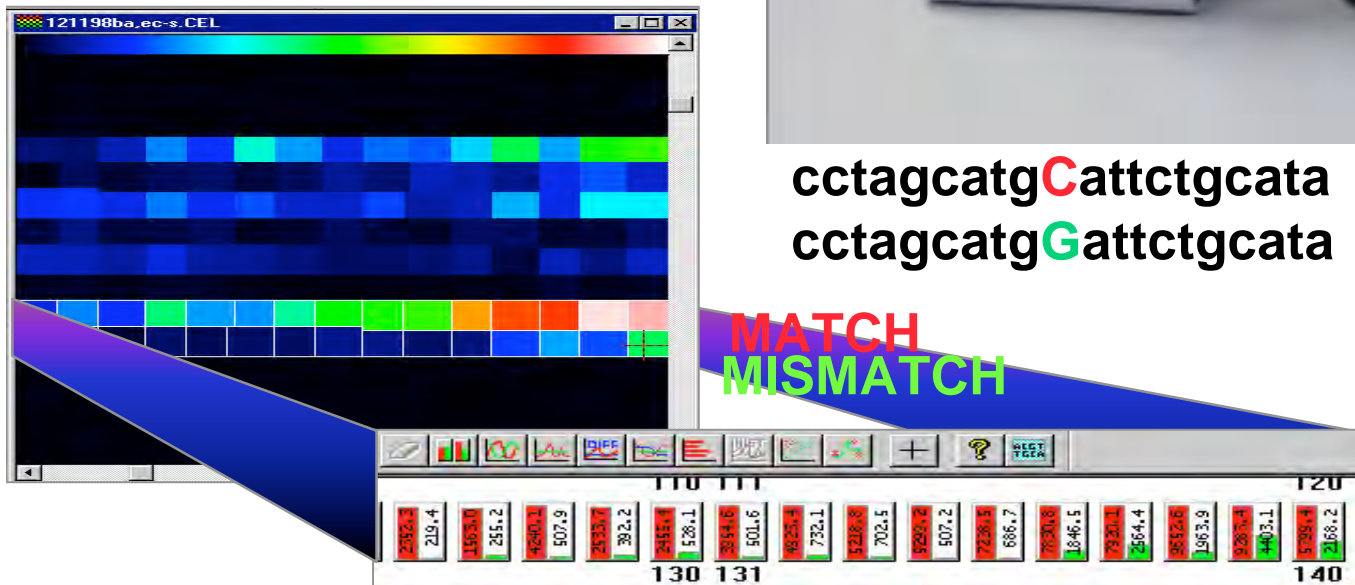
DOE 16s rDNA microarray

- Rapidly detect the composition and diversity of microbes in an environmental sample
- Massive parallelism - 550,000 probes in a 1.28 cm² array
- all 9,900 species in 16S rDNA database
- Single nucleotide mismatch resolution



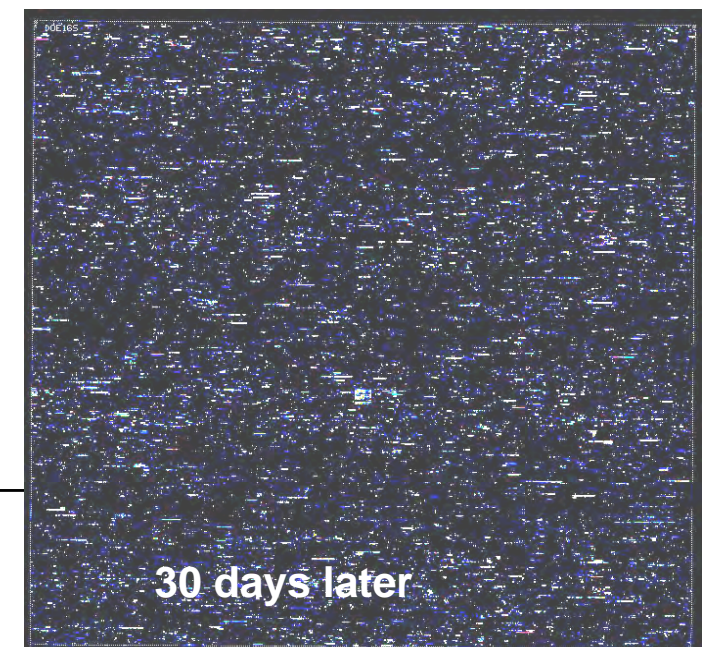
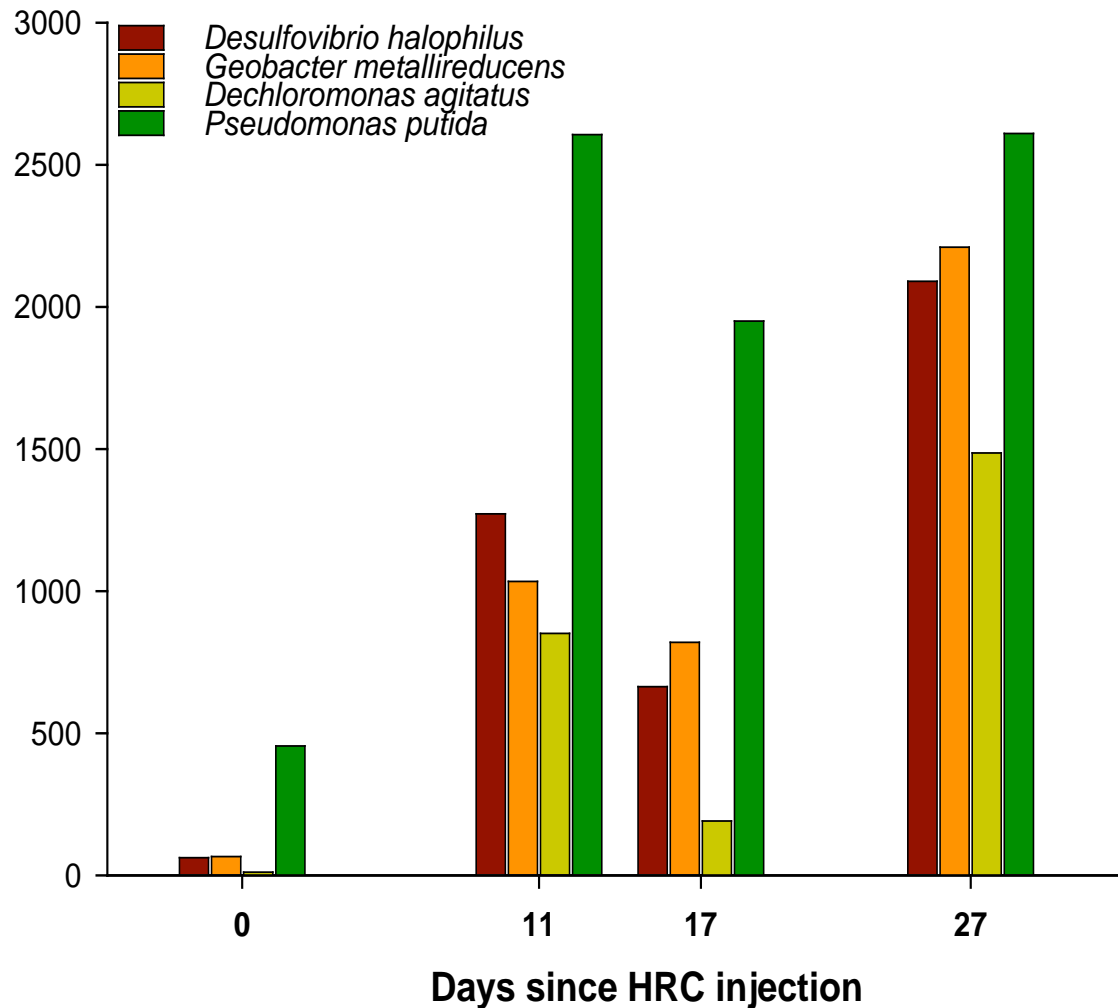
cctagcatgCattctgcata
cctagcatgGattctgcata

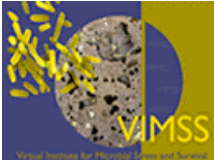
MATCH
MISMATCH



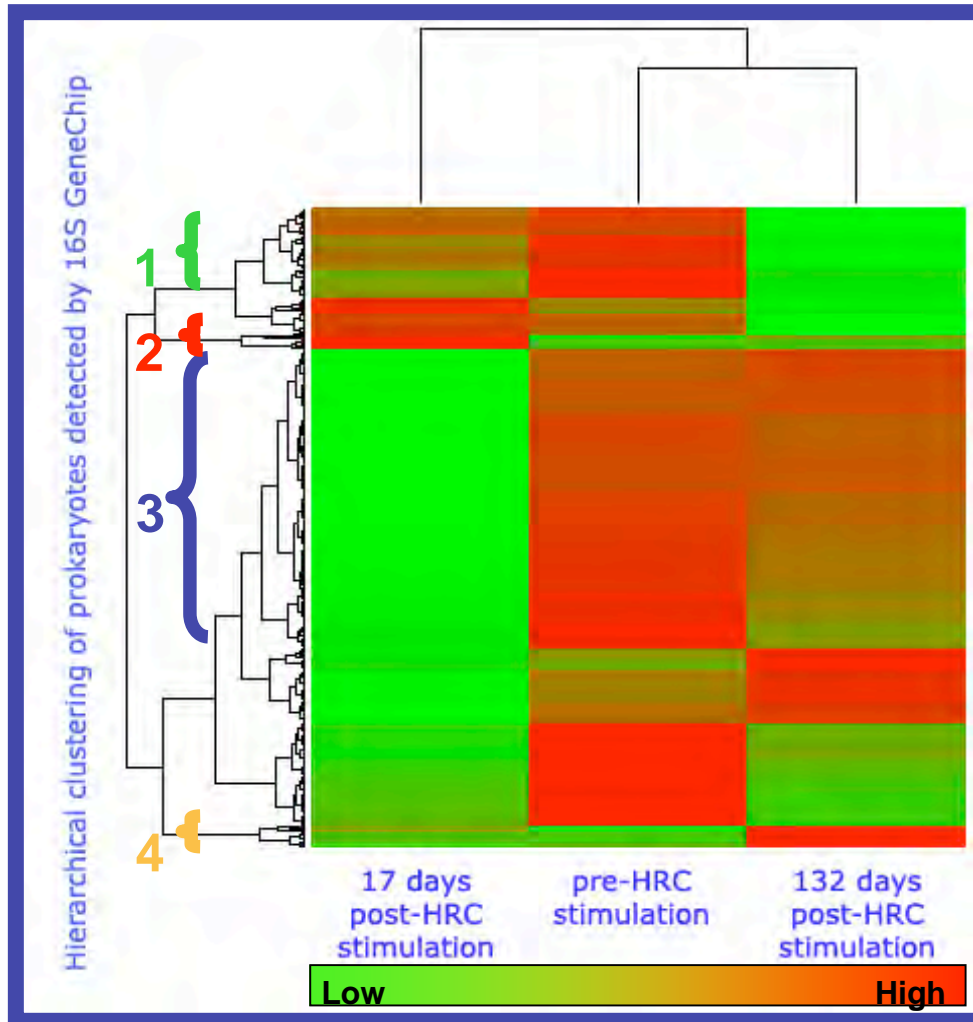
Microarray analysis of bacterial community changes during Cr(VI) remediation at Hanford 100H site:

Dynamics of some significant organisms.





High Density Microarray Analysis



Hierarchical clustering and heatmap plot of 16S GeneChip analysis of microbial community sub-families detected during chromate bioremediation. PCA groups are indicated by brackets.

Bacteria and Archaea Detected

Grouped according to response to HRC during chromate remediation

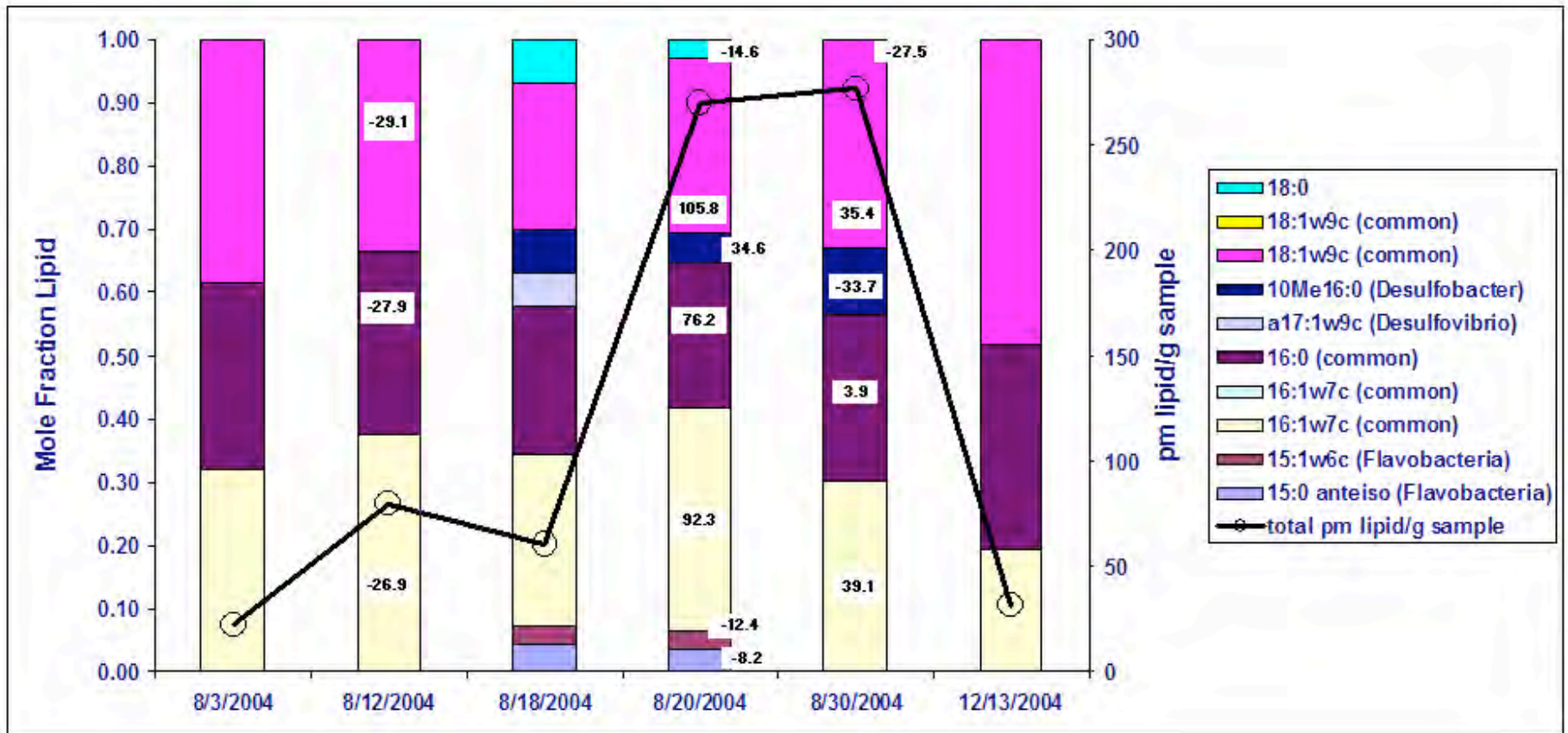
Group1 organisms decline
Pseudomonas, Burkholderia (Denitrifiers)
Acidithiobacillus, Thiothrix (Sulfur oxidizers)
Leptothrix (Iron oxidizer)

Group2 organisms increase then decline
Acidovorax, Thauera (denitrifiers)
Flavobacteria (aerobes, use glycerol)

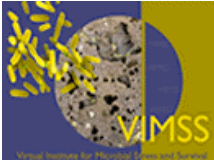
Group3 organisms decline then return
Mainly oligotrophic bacteria

Group 4 organisms increase in late stages
Legionella, Chlamydophila, Flectobacillus.

^{13}C Phospholipid Analysis

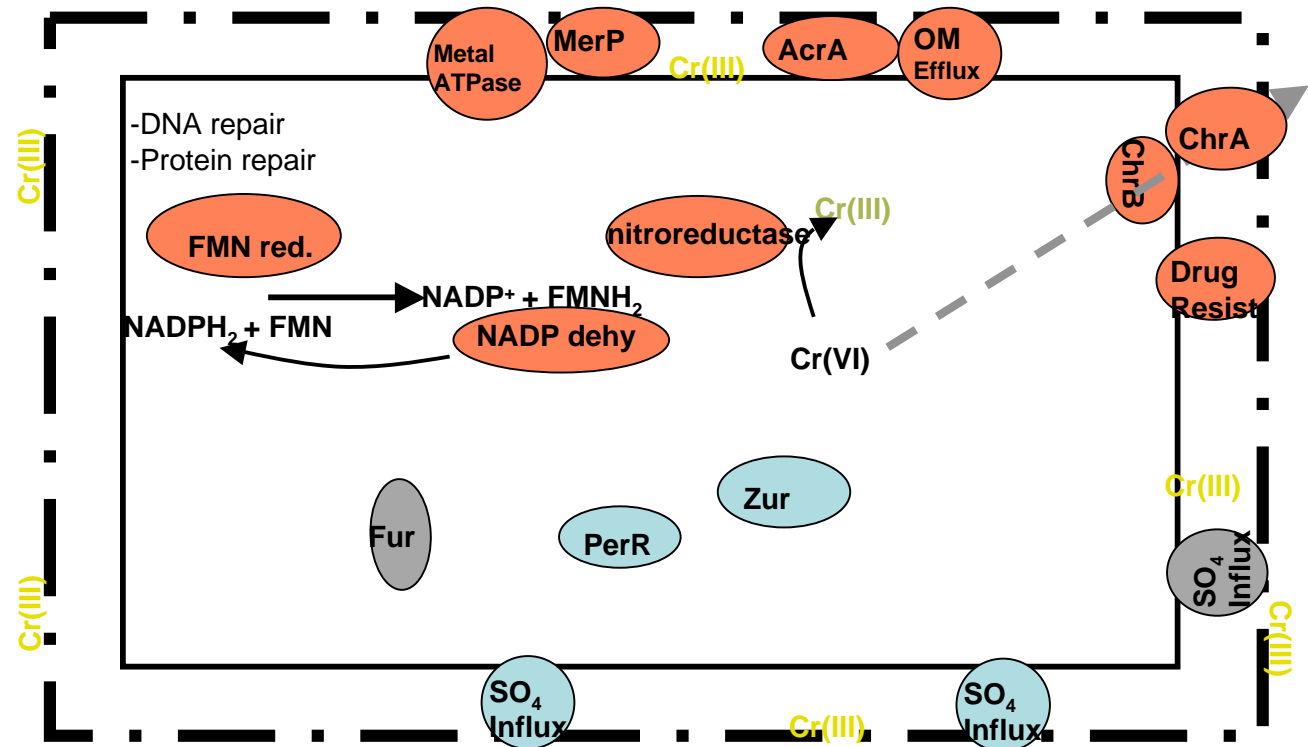


- General bacterial biomarkers indicate rapid enrichment in ^{13}C
- ^{13}C ratio is greater than expected (overall spiked HRC ratio was 15 per mil)
 - ^{13}C polylactate used as spike it is not esterified to glycerol backbone
 - it is released and consumed more rapidly
- Biomarkers for *Flavobacteriaceae* increased following injection but showed minimal enrichment with ^{13}C .
 - Flavobacteria* do NOT typically utilize lactate, but may use glycerol (backbone, unlabeled)

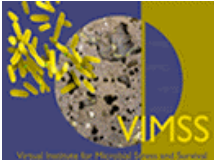


What Cellular Systems are Involved in Cr(VI) Responses in *Desulfovibrio vulgaris* Hildenborough?

- Sulfate influx down-expressed
- Metal efflux up-expressed
- *chrAB* up-expressed
- FMN dependent nitroreductase, NADH dehydrogenase, and FMN reductase up-expressed



Klonowska, A., He*, Z., He, Q., Hazen*, T.C., Thieman, S.B., Alm*, E.J., Arkin*, A.P., Wall*, J.D., Zhou*, J. and Fields*, M.W. Global Transcriptomic Analysis of Chromium(VI) Exposure of *Desulfovibrio vulgaris* Hildenborough Under Sulfate-Reducing Conditions. (in preparation)



Field Research Center, Oak Ridge TN

- Operated from 1951 to 1983
- 400 ft x 400 ft x 17 ft deep
- Over 2.5 million gallons waste/year
- Wastes contained nitrate, uranium, Tc-99, metals, VOCs, high TDS, and low pH (<2.0)
- Neutralized in 1984 capped in 1988

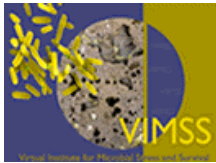


Currently a parking lot

S-3 Disposal Ponds During Denitrification

S-3 Waste Disposal Ponds

- Unlined
 - Received liquid nitric acid and uranium-bearing wastes
- ~ 320 million liters 1951-1983



Environmental Whole-Genome Amplification To Access Microbial Populations in Contaminated Sediments

- Recovery of adequate amounts of DNA for molecular analyses can often be challenging in stressed microbial environments.
- Developed multiple displacement amplification (MDA) methods for unbiased, isothermal, amplification of DNA
- Subsequently applied these technologies to understand stressed, low biomass, populations in multiple sediments contaminated with Uranium on the Oak Ridge Reservation
- Over 4000 clones were end sequenced. 5% of all clones were identified as belonging to Deltaproteobacteria (primarily, *Geobacter* and *Desulfovibrio*-like)
- Significant overabundance of proteins (COGs) associated with: 1) Carbohydrate transport & metabol. 2) Energy production & conversion, 3) Posttranslational modification, protein turnover, & chaperones. --- All of which may be important in adaptation to environmental stressors such as low pH, high contaminate loads, and oligotrophic nature of the subsurface environment

Statistics on amplified metagenome library end-sequences

Library	Area 3, Deep		Area 3, Shallow		Area 2		Total	%
	Deep	%	Shallow	%		%		
Number of clones sequenced	960		864		864			
Sequences generated	1,920		1,728		1,728			
Quality sequences ^a	1,394	100	1,118	100	1,509	100	4,021	100
Sequences that form contigs	370	26.5	152	13.6	141	9.3	663	16.5
Number of contigs assembled	101		53		54		208	
Sequences with similarities to known proteins ^b	928	66.6	692	61.9	990	65.6	2,610	64.9
Highest similarity to bacterial proteins	901	64.6	629	56.3	890	59.0	2,420	60.2
Highest similarity to Deltaproteobacteria proteins	35	2.5	23	2.1	155	10.3	213	5.3
Highest similarity to archaeal proteins	12	0.9	43	3.8	79	5.2	134	3.3
Highest similarity to eukaryotic proteins	12	0.9	18	1.6	21	1.4	51	1.3

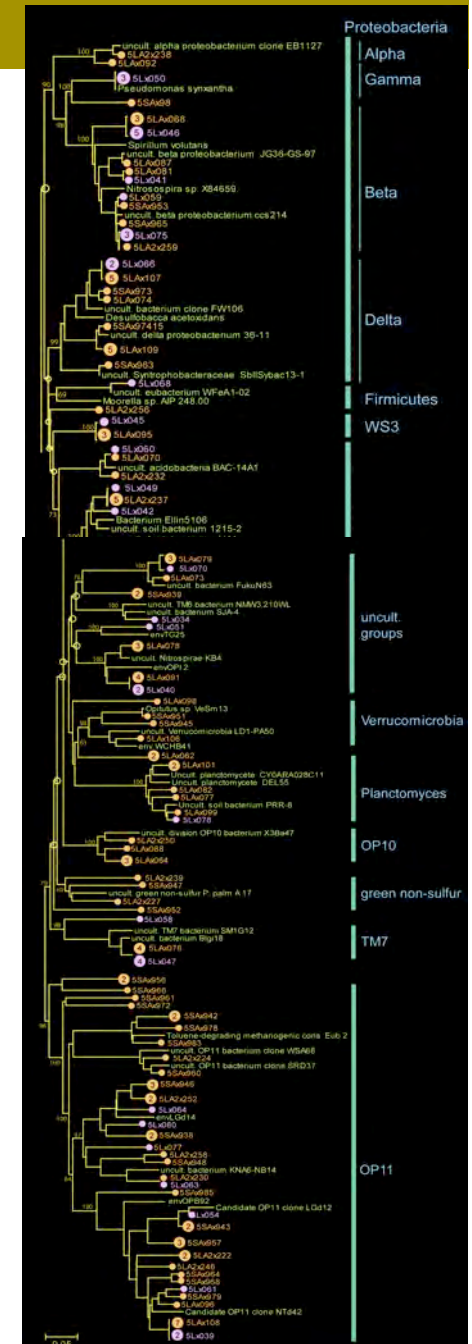
a. Sequences >400nt in length

b. e-values <1e-10 from BLASTX searches against the NCBI protein database

Abulencia, C.B., Wyborski, D.L., Garcia, J., Podar, M., Chen, W., Chang, S. H., Chang, H.W., Watson, D., Brodie, E.L., Hazen, T.C. and Keller, M. (2006)

Environmental Whole-Genome Amplification to Access Microbial Populations in Contaminated Sediments. *Appl. Environ. Microbiol.* **72**(5):3291-3301 [[download](#)

[pdf](#)]

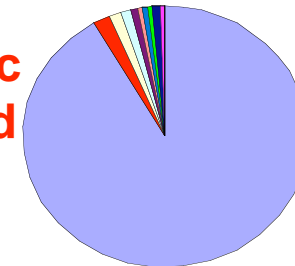
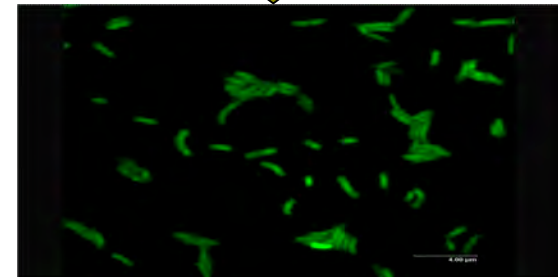


Metagenomic Analysis of NABIR FRC Groundwater Community

Data: Jizhong Zhou et al.

Metagenomic sequencing:

- Almost like a mono-culture
- 52.44 Mb raw data assembled into contigs totaling ~5.5 Mb
- 224 scaffolds (largest 2.4 Mb)
- Genes important to the survival and life style in such environment were found



■	Frateuria	99%
■	Herbaspirillum	99%
□	Alcaligenes	98%
□	Frateuria	100%
■	Frateuria	96%
■	Frateuria	95%
■	Burkholderia	99%
■	Frateuria	96%
■	Burkholderia	99%
■	Frateuria	98%

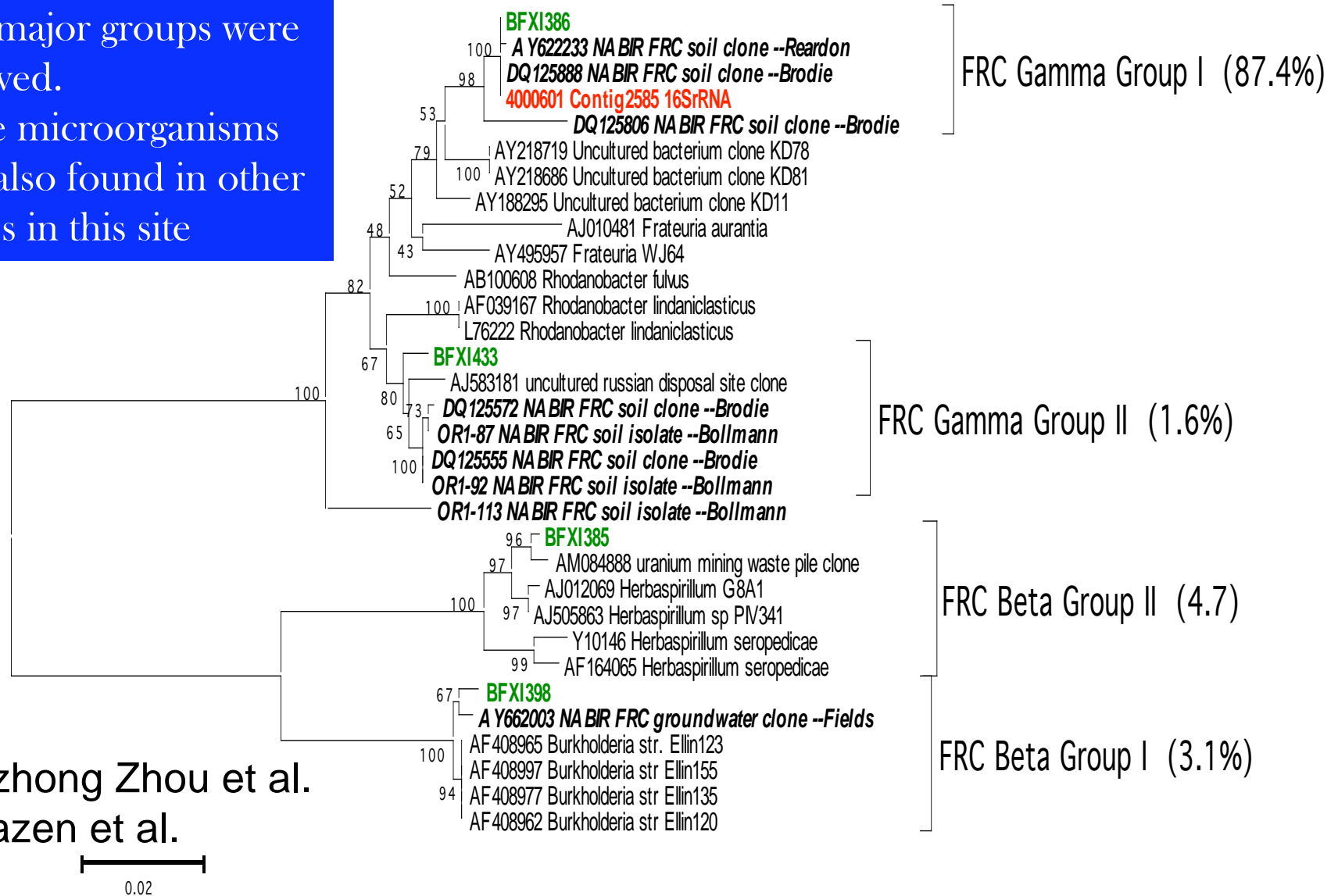
Extremely low diversity:

- Dominated by *Frateuria*-like organism
- At least 2 *Frateuria* phylotypes
- Azoarcus species: less abundant

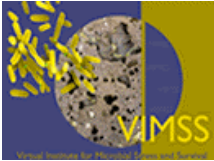
These results suggest that contaminants have dramatic effects on the groundwater microbial communities, and these populations are well adapted to such environments.

Phylogenetic Tree of SSU rRNA Genes

- Four major groups were observed.
- These microorganisms were also found in other studies in this site

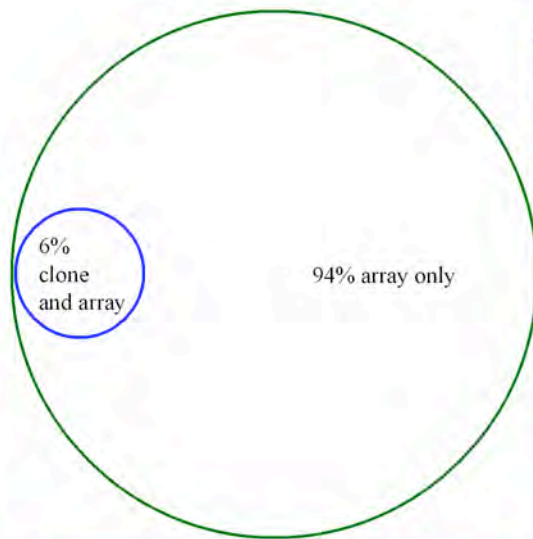


Data: Jizhong Zhou et al.
Terry Hazen et al.



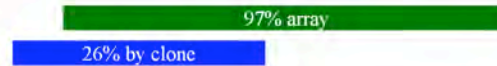
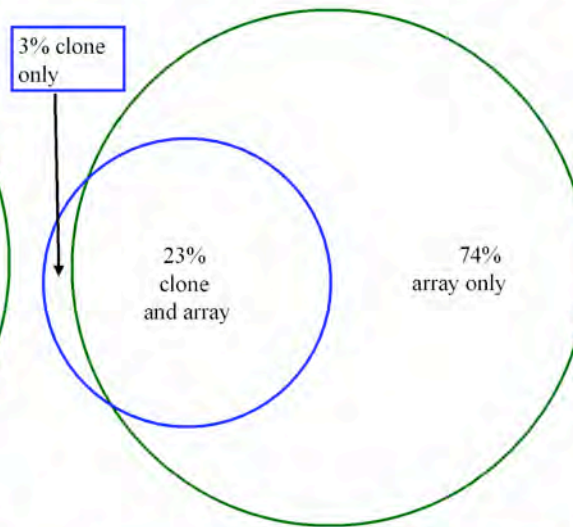
Accuracy V Clone libraries

Cr(VI) groundwater



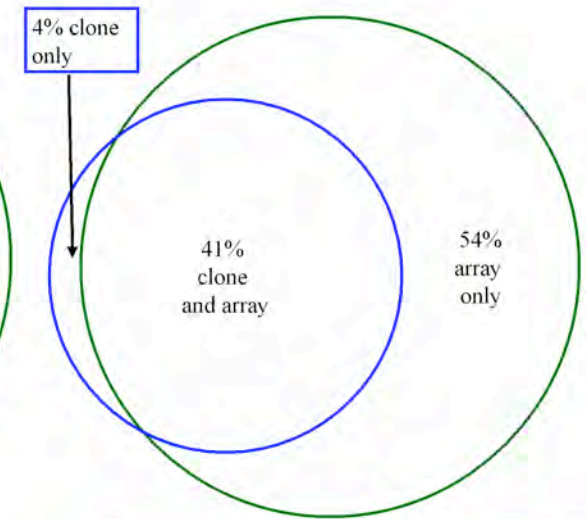
384 clones

U(VI) subsurface soil

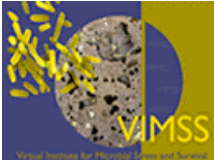


768 clones

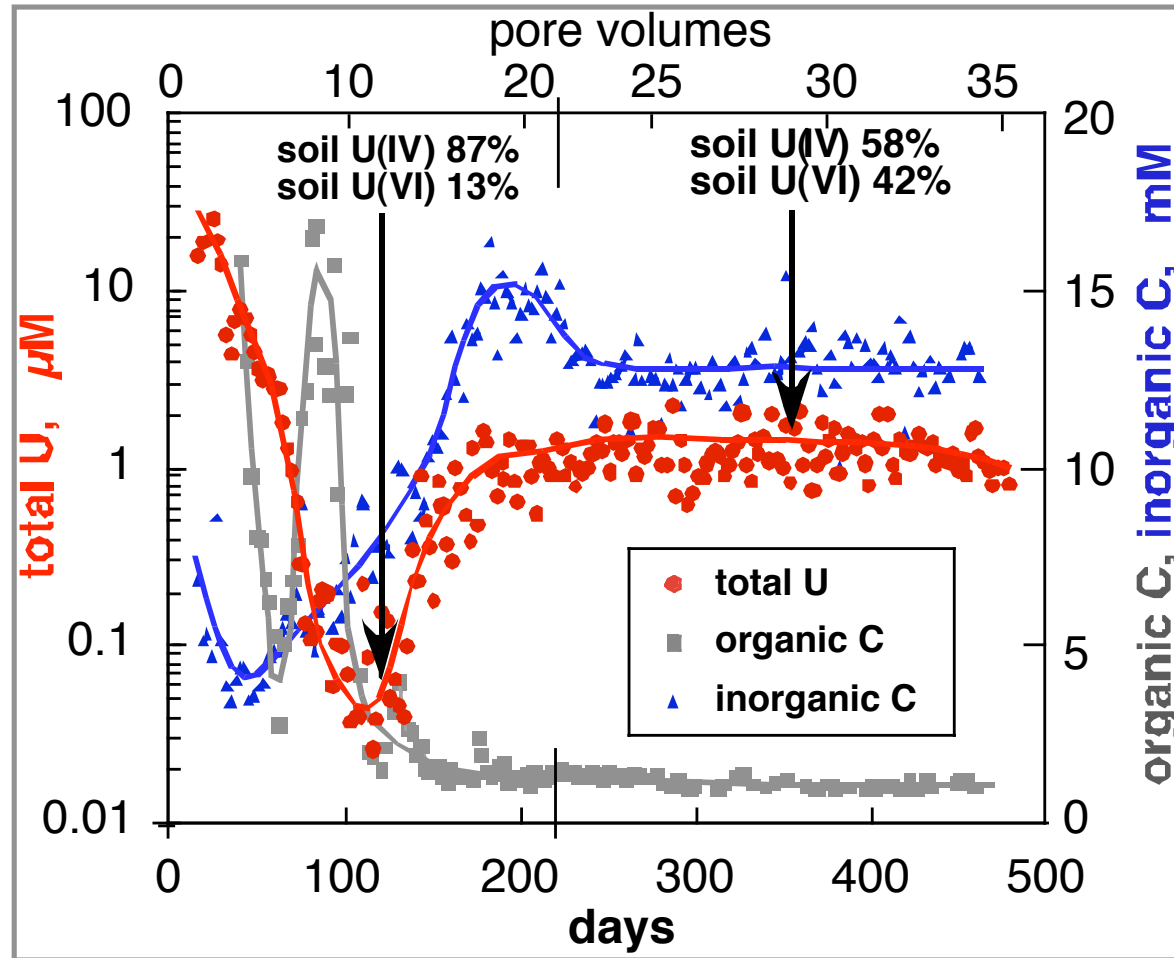
Urban aerosol



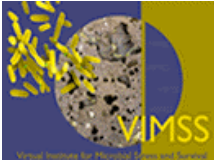
768 clones



Uranium Anaerobic Reoxidation

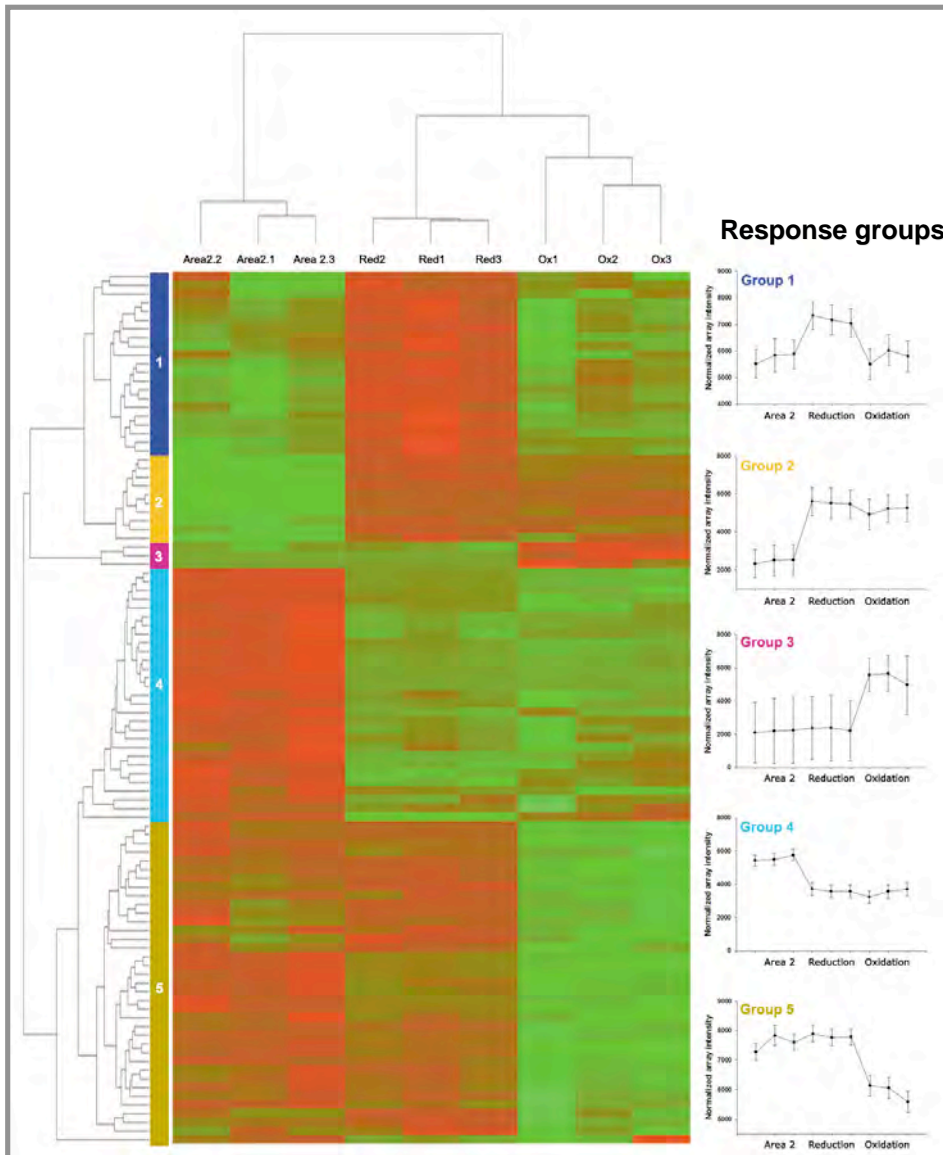


Microbial metabolism - CO_2 produced increasing dissolved IC



Bi-directional clustering of array data

Bacterial taxa



Column sediment samples

Major components of response groups

- *Arthrobacter sp.*
- Alpha-proteobacteria
- *Azoarcus sp.*

- *Geothrix*
- *Geobacter*
- *Anaeromyxobacter*

- *Acidobacteria*
- *Desulfovibrio*

- *Actinobacteria*
- *Firmicutes*
- *Alpha-proteobacteria*

- *Actinobacteria*
- *Alpha-proteobacteria*

Uranium
reducers

Syntrophy?

Brodie, E. L., T. Z. DeSantis, D. C. Joyner, S. M. Baek, J. T. Larsen, G. L. Andersen, T. C. Hazen, P. M. Richardson, D. J. Herman, T. K. Tokunaga, J. M. Wan, and M. K. Firestone. 2006. Bacterial population dynamics during uranium reduction and re-oxidation: Application of a novel high density oligonucleotide microarray approach. *Appl. Environ. Microbiol.* 72:6288-6298

Integrated approach to the Phytoremediation of Lead-Contaminated Lands Katowice, Poland

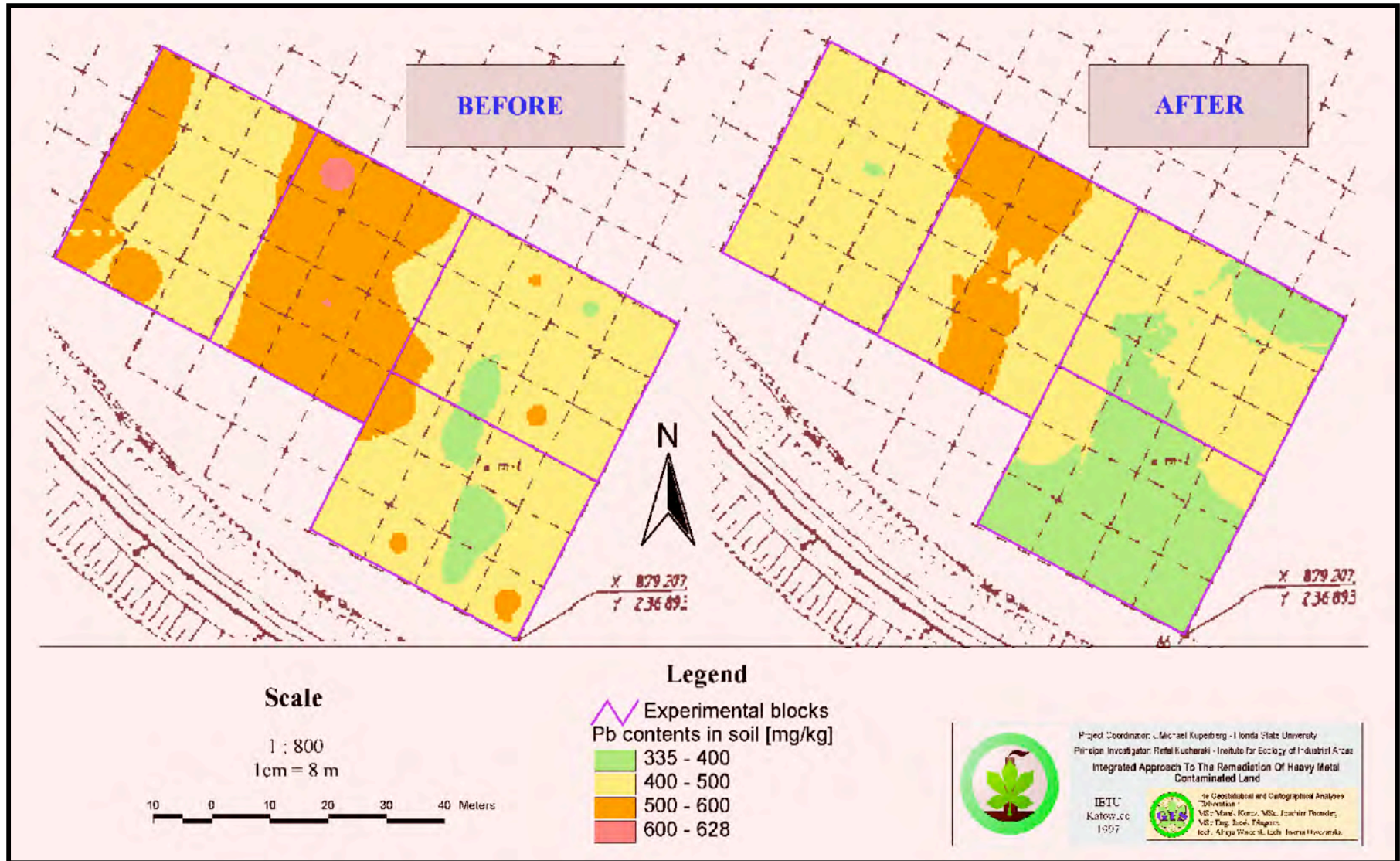


- Site selection & characterization
- Screening for best heavy metal accumulators
- Selection of amendments
- Amendment application technique
- Harvest and biomass disposal
- Ecological risk assessment
- Economic evaluation



U.S. DOE Office of Environmental Management
Institute for Ecology of Industrial Areas (Katowice, Poland)
Florida State University, Central European Advanced Technologies
Edenspace (Phytotech)

Results after one year





Summary

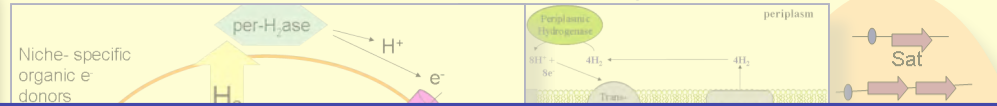
- Bioremediation holds great promise for remediating some of our most recalcitrant contaminated sites (oil, chlorinated solvents, Pb, U, Cr). Biostimulation and natural attenuation are the most dominant field applications to date. **Good**
- Critical biogeochemical characteristics and monitoring need to be considered for selection of the most appropriate strategy and monitoring (factors: sensitivity, specificity, quantification, sorption, reoxidation, mobilization, toxicity, bystander effects), bioaugmentation has been very limited and may always be). **Bad**
- Phytoremediation and bioaccumulation/biosorption strategies also show promise but may have fatal flaws (life cycle analyses that include all risks and cost are critical). **Ugly**
- Biomobilization and treatment trains that end in natural attenuation maybe the best long-term solutions especially for mixed waste.
- A Systems Biology approach may be one of the only ways that we that we can enable sustainable environmental biotechnology applications.

Virtual Institute of Microbial Stress and Survival

Environmental Monitoring



Pathway Models



Pathway Inference

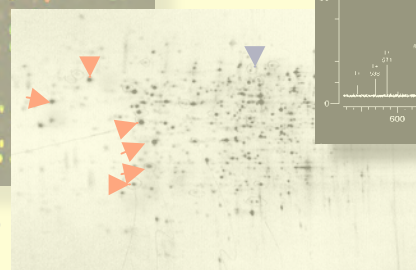
Overall VIMSS Goals

- To understand the mechanisms by which microbes adapt and survive
- To elucidate how they carry out mission critical processes
- To rapidly characterize new microbes to the level of a model microbe

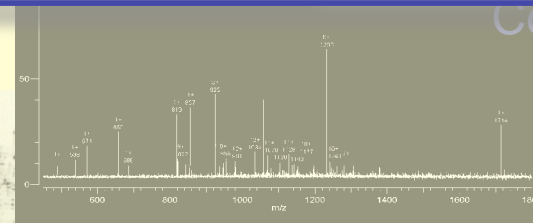
sFTIR
In situ
physiology



DNA Microarrays

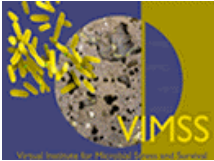


Proteomics



Metabolomics

Comparative Genomics



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Adam Arkin, Eric Alm, Kat Huang, Dylan Chivian, Janet Jacobson, Jay Keasling, Aindrila Mukhopadhyay, Eoin Brodie, Sharon Borglin, Hoi-Ying Holman, Jil Geller, Elenor Woezi, Jenny Lin, Dominique Joyner, Rick Huang, Romy Chakraborty, Boris Faybishenko, Mark Conrad, Joern Larsen, Zouping Zheng, Gary Andersen, Todd DeSantis, Tetsu Tokunaga, Jiamin Wan, Susan Hubbard, Ken Williams, John Peterson, Natalie Katz, Jill Banfield, Tamas Torok, Seung Baek, Don Herman, Mary Firestone



Phil Long, et al.



Steve Koenigsberg, Ana Willet



Paul Richardson, Phil Hugenholtz



Judy Wall, et. al.



Mathew Fields, et. al.



Stephen Sutton, Matthew Newville



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Massachusetts Institute of Technology

Eric Alm



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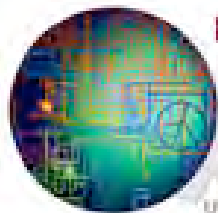


Indiana University

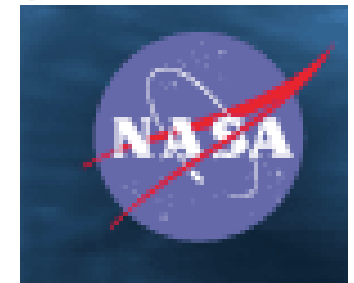
Lisa Pratt, et. al.

Princeton

T. C. Onstott, et. al.



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Center for Environmental Biotechnology

<http://www-esd.lbl.gov/CEB>

Virtual Institute for Microbial Stress and Survival

<http://vimss.lbl.gov>

<http://www.microbesonline.org> - Comparative Genomics Database, Genome Browser, Operon Browser, Regulon Browser, Metabolic Maps of >430 sequenced bacteria and archaea

<http://greengenes.lbl.gov> - 16S rRNA gene database and workbench

Environmental Remediation Technology Program

<http://www-esd.lbl.gov/ERT>

Ecology Department

<http://www-esd.lbl.gov/ECO>

DOE Environmental Remediation Sciences Program

<http://www.lbl.gov/ERSP>

DOE Genomics:GTL Program

<http://doegenomestolife.org>

NASA Indiana Princeton Tennessee Astrobiology Initiative (IPTAI)

<http://www.indiana.edu/%7Edeephlife/homepg.html>

